

BMJ Open is committed to open peer review. As part of this commitment we make the peer review history of every article we publish publicly available.

When an article is published we post the peer reviewers' comments and the authors' responses online. We also post the versions of the paper that were used during peer review. These are the versions that the peer review comments apply to.

The versions of the paper that follow are the versions that were submitted during the peer review process. They are not the versions of record or the final published versions. They should not be cited or distributed as the published version of this manuscript.

BMJ Open is an open access journal and the full, final, typeset and author-corrected version of record of the manuscript is available on our site with no access controls, subscription charges or pay-per-view fees (http://bmjopen.bmj.com).

If you have any questions on BMJ Open's open peer review process please email info.bmjopen@bmj.com

BMJ Open

Fabry disease due to D313Y and novel GLA mutations

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-017098
Article Type:	Research
Date Submitted by the Author:	23-Apr-2017
Complete List of Authors:	Koulousios, Konstantinos; University of Thessaly Faculty of Medicine, Stylianou, Konstantinos; University Hospital of Heraklion Pateinakis, Panagiotis; Geniko Nosokomeio Thessalonikis Papageorgiou Zamanakou, Maria; CeMIA SA Loules, Gedeon; CeMIA SA Manou, Eleni; Geniko Nosokomeio Thessalonikis Papageorgiou Kyriklidou, Parthena; Geniko Nosokomeio Thessalonikis Papageorgiou Katsinas, Christos; Mpodosakeio General Hospital Ouzouni, Alexandra; Geniko Nosokomeio Kabalas Kyriazis, John; Geniko Nosokomeio Chiou Skylitseio Speletas, Matthaios; University of Thessaly Faculty of Medicine Germenis, Anastasios E.; University of Thessaly Faculty of Medicine
Primary Subject Heading :	Genetics and genomics
Secondary Subject Heading:	Renal medicine, Cardiovascular medicine, Neurology, Diagnostics
Keywords:	Fabry disease, D313Y GLA mutation, Novel GLA mutations, Kidney biopsy, Misdiagnosis

SCHOLARONE™ Manuscripts

1 Fabry disease due to D313Y and novel GLA mutations

- 2 Konstantinos Koulousios^{a*}, Konstantinos Stylianou^b, Panagiotis Pateinakis^c, Maria
- 3 Zamanakou^d, Gedeon Loules^d, Eleni Manou^c, Parthena Kyriklidou^c, Christos Katsinas^e,
- 4 Alexandra Ouzouni^f, John Kyriazis^g, Matthaios Speletas^a, Anastasios E. Germenis^a
- 5 *Corresponding author
- 6 ^a Department of Immunology & Histocompatibility, School of Medicine, University of
- 7 Thessaly, Panepistimiou 3, GR-41500 Biopolis, Larissa, Greece
- 8 b University Hospital of Heraklion, Heraklion, Greece
- 9 ^c Papageorgiou General Hospital, Thessaloniki, Greece
- 10 d CeMIA SA, Makriyianni 31, GR-41334 Larissa, Greece
- 11 ^e Mpodosakeio General Hospital, Ptolemaida, Greece
- 12 ^f Kavala General Hospital, Kavala, Greece
- 13 ^g Chios General Hospital, Chios, Greece
- 14 Konstantinos Koulousios (konkoul@gmail.com)
- 15 Konstantinos Stylianou (kstylianu@gmail.com)
- 16 Panagiotis Pateinakis (pateinakis@hotmail.com)
- 17 Maria Zamanakou (mzamanakou@cemia.eu)
- 18 Gedeon Loules (gedloules@cemia.eu)
- 19 Eleni Manou (elmanou@webcoreforscience.com)
- 20 Parthena Kyriklidou (renakiri@yahoo.it)
- 21 Christos Katsinas (katsinas@otenet.gr)
- 22 Alexandra Ouzouni (alouzouni2808@gmail.com)
- John Kyriazis (jks@otenet.gr)
- 24 Matthaios Speletas (maspel@med.uth.gr)



27	Abstract
28	Objectives: Our aim is to report five novel <i>GLA</i> mutations resulting in FD and provide
29	evidence of pathogenicity of the D313Y mutation regarding which contradictory data have
30	been presented in the literature.
31	Setting and participants: 25 family members of nine unrelated patients with definite FD
32	diagnosis, ten clinically suspected cases and eighteen members of their families were included
33	in this polycentric cohort study.
34	Primary and secondary outcome measures: Genotyping and measurement of lyso-Gb ₃ was
35	performed in all individuals. The $\alpha\text{-Gal}\ A$ activity was measured in all males as well as
36	plasma and urine Gb ₃ concentration in selected cases. Optical and electron microscopy was
37	performed in kidney biopsies of selected patients. All the above were evaluated in parallel
38	with the clinical data of the patients.
39	Results: Sixteen new cases of FD were recognised, three of which were carrying already
40	described GLA mutations. Five novel GLA mutations, namely c.835C>T, c.280T>A,
41	c.924A>C, c.511G>A and c.453C>G, resulting in a classical FD phenotype were identified.
42	Moreover, FD was definitely diagnosed in five patients carrying the D313Y mutation. Eight
43	D313Y carriers were presenting sings of FD despite not fulfilling the criteria of the disease,
44	two had no FD signs and two others were apparently healthy.
45	Conclusions: Five novel GLA pathogenic mutations are reported and evidence of
46	pathogenicity of the D313Y mutation is provided. It seems that the D313Y mutation is
47	related with a later-onset milder than the typical phenotype with normal lysoGb ₃
48	concentration. Our study underlines the significance of family members genotyping and
49	newborn screening in avoiding misdiagnoses and crucial delays of diagnosis and treatment of
50	the disease.

Strengths and limitations of this study

- Novel GLA mutations resulting to a classical Fabry disease phenotype are presented.
- The clinical impact of the D313Y mutation of the GLA mutation is analysed in a significant number of male and female carriers at various ages.
- This study offers strong evidence that the D313Y mutation could be pathogenic, indicating that therapy should be considered when appropriate.
- The main limitation is the lack of detailed clinical data in older participants.

INTRODUCTION

FD or Anderson-Fabry disease is an X-linked inherited metabolic disorder, that results from mutations in the α -gal A gene (*GLA* gene), leading to reduction of the enzyme activity and subsequent accumulation of Gb₃ (or GL-3) in plasma, urine and cellular lysosomes throughout the body. These depositions cause a multisystemic pathology with life-threatening manifestations, including renal failure, cardiac and cerebrovascular disease [1, 2].

More than 600 currently known *GLA* mutations have been identified [3, 4], as causing a variety of clinical manifestations. Most of them are unique to a family (private) and therefore genotype-phenotype correlation is limited [5]. Diagnosing FD is challenging due to the range of disorders that mimic the disease and the great variety of atypical clinical presentations. As a result underdiagnosis and misdiagnosis of FD lead to late diagnosis, delays in appropriate treatment and a subsequent negative prognosis [6]. Human genetic analysis must be performed, in order to exclude or verify a mutation of the *GLA* [7]. Once a diagnosis has been made, biochemical and molecular genetic analysis, as well as genetic counselling, should be made available to all family members [8]. A detailed pedigree analysis for each patient presenting with FD is crucial [9], as it can inform the diagnosis of the proband and the identification of all at-risk relatives [10].

Our aim is to report five novel *GLA* mutations resulting in FD and provide evidence of pathogenicity of the D313Y mutation regarding which contradictory data have been presented in the literature.

METHODS

Study population

A cohort of 62 subjects was involved in the study selected between families of 9 unrelated patients with definite diagnosis of FD as well as amongst cases with nephrological, cardiac or neurological symptoms suspicious of this diagnosis. Eighteen family members of the last cases were also examined after the confirmation of diagnosis. The presence of the D313Y mutation in Greek population was examined by genotyping of 145 apparently healthy subjects (70 males, 75 females).

Written informed consent was obtained from each subject or an accompanying relative, where legally appropriate. The study was approved by the institutional review board of the University of Thessaly, Larissa.

Clinical assessment

Patients' medical records were revaluated and a detailed medical history of the family members was obtained especially in regard with heart or kidney disease, cerebrovascular events, death at young age and respective causes of death. All study participants underwent physical examination particularly focused on cardiac, renal and neurological signs and symptoms. A detailed pedigree was constructed for every family and newborn screening was performed once.

Laboratory evaluation

In DBS, we measured α -Gal A activity in all male subjects by tandem mass spectrometry, lyso-Gb₃ in all subjects by HPLC and tandem spectrometry [11] as well as plasma and urine Gb₃ concentration in selected cases by tandem mass spectrometry.

Optical and electron microscopy was performed for the study of kidney biopsies occasionally.

Genotyping

Analysis of primary data was conducted with Ion Reporter software v.5.2 (Thermo Scientific). Common polymorphisms (UCSC Common SNPs) were excluded and pathogenicity of variations was predicted by bioinformatic analysis using PhyloP, SIFT, Grantham and PolyPhen tools, in comparison to their global (1000 Genomes Global Minor Allele Frequency, ExAC) and European frequency (5000 Exomes European Minor Allele Frequency). The characterization of variants was based on the recommendations of the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology [12]. Novel mutations were verified by standard Sanger sequencing using Variant Reporter software v1.1 (Applied Biosystems).

RESULTS

Six (all with definite FD) out of the 62 genotyped subjects were carrying four previously described *GLA* mutations: c.334C>T (p.Arg112Cys, R112C), c.644A>G (p.Asn215Ser, N215S), c.1153A>C (p.Thr385Pro, T385P) and c.782G>T (p.Gly261Val, G261V).

The c.937G>T (p.Asp313Tyr, D313Y, NM_000169.2) mutation was revealed in seventeen individuals (54±14, range 27–78 years), seven males (61±11, range 45–78 years) and ten females (49±15, range 27–70 years) but in none of the healthy subjects. Patients' clinical and laboratory findings are presented in Table 1.

The diagnosis of FD was definitely posed according to the recently published criteria of the disease [13], in five carriers of this mutation (54 \pm 9, range 45–65 years). The first of them (patient no.1, Table 1), a 52-year-old man was initially diagnosed at the age of 48 with CKD stage III and was not presenting cardiac or other FD signs. Kidney biopsy performed because of non-nephrotic proteinuria, microscopic haematuria and raised serum creatinine, revealed focal and segmental glomerulosclerosis (FSGS) – collapsing variant. At that time enzyme activity and plasma lyso-Gb3 concentration were normal. Three years after initial presentation the patient was suffering end stage renal disease and extreme acroparesthesias. Revaluation of kidney biopsy by higher magnification uncovered focal cytoplasmic microvacuolization of enlarged podocytes (Fig.1) while a decreased by 50% α -gal A activity and an increased plasma (7.52 nmol/mL, reference: 0.8-4.52) and urine Gb3 concentration (147.49 nmol/g, reference: <29.00) were detected. The patient commenced dialysis and ERT with rapid clinical improvement.

A second carrier of the D313Y mutation was a 46-year-old female (patient no. 4, Table 1) who had suffered a TIA and two ischaemic strokes that had been considered of unknown origin. On revaluation, one year after the last stroke the patient appeared with microalbuminuria, oedema and acroparesthesias on both hands. The microalbuminuria was duplicated after three months and acroparesthesias worsened. Gb₃ concentration was pathological in urine (54.08 nmol/g) but normal in plasma. LysoGb₃ concentration was normal at that time and remained stable for one more year.

The mother of the above patient (patient no. 5, Table 1), 65 years old, had been diagnosed at the age of 50 with MS and was receiving relative medication, without clinical benefit. She was also receiving treatment for pain in extremities that were attributed to RA. During the recent years the patient suffered mobility impairment (reported as spastic quadriplegia after a neurological examination), depression and dementia. The evaluation of

the patient revealed pathological plasma Gb₃ concentration (4.7 nmol/mL), WML on brain MRI and normal kidney function. LysoGb₃ concentration was normal and remained so during a 6-month-follow up.

Another carrier of the D313Y mutation was a 45-year-old male (patient no.7, Table 1) on dialysis due to CKD by the age of 25. No kidney biopsy was performed at that time. He reports episodes of haematuria during childhood and adolescence, attributed at that time to vesicoureteral reflux. Enzyme activity was slightly decreased (2.4 µmol/l/h, reference: ≥2.6) and plasma lyso-Gb₃ concentration was normal. Brain MRI revealed WML and vertebrobasilar vessel changes. Moreover, increased echogenicity of cardiac interventricular septum on the echocardiogram and sensorineural hearing loss of higher frequencies. The patient commenced ERT.

The last patient with D313Y mutation definitely diagnosed as suffering FD was a 60-year-old female (patient no.17, Table 1) who was presenting cornea verticillata corneopathy, WML and ischaemic infarcts on brain MRI despite that no stroke is reported, acroparesthesias and GI symptoms (pain-diarrhoea) since adolescence, hypohidrosis, hearing loss and LVH.

Among the remaining twelve D313Y mutation carriers there were two patients with no FD signs and eight patients (55 ± 17 , range 27–78 years) presenting other FD signs mainly neurological that, however, can not document a definite diagnosis of FD. Especially patient no.6, Table 1 (62-year-old female) suffers acroparesthesias and GI symptoms since adolescence and presents with lysoGb₃ at 1.7 ng/ml (reference: ≥ 1.8). Accordingly patient no.16, Table 1 (64-year-old male) presents with CKD, diabetes mellitus and hearing loss, while on brain MRI presents multiple ischaemic infracts despite that no stroke is reported. The remaining two D313Y mutation carriers are apparently healthy.

Novel GLA mutations (5) (Fig. 2) were detected in sixteen members (42±19, range 1– 73 years) of five unrelated families (Table 2), all fulfilling the diagnostic criteria of a definite diagnosis of FD [13]. The c.835C>T mutation (p.Gln279Ter, Q279X) in exon 6 of the GLA gene was identified in four members (30±24, range 1–60 years) of a Greek family. The proband, a 31-year-old male, was presented at the age of 23 with proteinuria (3.5 gr/24h), microscopic haematuria, slightly deteriorated kidney function, right bundle brunch block, mild hypertension and angiokeratomas in the arms and the loin area. He reported pain in the extremities especially during infections and inability to sweat. The α-gal A activity was close to zero. Cardiac MRI showed moderate LVH. The kidney biopsy showed cytoplasmic vacuolization and extended lysosomal accumulations in all types of kidney cells, especially in the podocytes (Fig. 3). The patient is currently under ERT with beneficial results in regard with kidney function, proteinuria and pain. Three other members of the family were revealed having with the same mutation and all presented clinical signs of FD. The proband's mother (60 years old) reported a possible TIA at the age of 32. Kidney examination showed albuminuria (> 500mg/24h), microscopic haematuria and normal kidney function. Cardiac MRI revealed severe LVH (cardiac interventricular septum over 19mm) and on skin examination she showed angiokeratomas in the arms. Her mother (the proband's grandmother) had died at the age of 62, due to cardiac arrest. She suffered from severe LVH and acroparesthesias, which at that time were attributed to Raynaud's phenomenon. The proband's sister (27-year-old) has elevated lyso-Gb₃ concentration (2.7 ng/ml) and screening of her newborn daughter revealed the mutation too.

Four patients (46±11, range 30–56 years) belonging to another Greek family were carrying the c.280T>A (p.Cys94Ser, C94S) mutation. The proband, a 48-year-old female, was diagnosed with FD a year before after presenting increased lyso-Gb₃ concentration (5.4 ng/ml), increased plasma Gb₃ concentration (6.14 nmol/ml), microalbuminuria of no other

origin, cornea verticillata corneopathy, acroparesthesias in both hands and dyshidrosis.

Clinical and laboratory data of the other suffering members of the family are presented in Table 2.

The c.924A>C mutation (K308N - p.Lys308Asn) in exon 6 of the *GLA* gene was identified in three members (44±26, range 17–69 years) of a Greek family. The proband, a 46-year-old male, was presented at the age of 34 with albuminuria (0,5-0,6 gr/24h), microscopic haematuria since ten years and slight hypertension. The kidney function was normal. Kidney biopsy showed slight mesangial proliferative damages and cytoplasmic microvacuolization of podocytes. The α -gal A activity was almost zero. After 10-year-follow up he suffered of LVH and proteinuria (1.8 gr/24h). The patient is currently under ERT. His mother and daughter carry the same mutation. The probands grandmother had died at the age of 68, suffering of severe LVH and end-stage heart failure.

The c.511G>A mutation (G171S - p.Gly171Ser) in exon 3 of the *GLA* gene was identified in two members (37 \pm 4, range 34–39 years) of an Albanian family living in Greece. The proband, a 39-year-old male, was diagnosed at the age of 32 with severely deteriorated kidney function and proteinuria. No biopsy was performed at that time, due to the small size of the kidneys. After nearly a year he presented with severe clinical and laboratory findings of acute renal failure and need of dialysis. Normal kidney function was never restored. FD was definitely diagnosed at the age of 37, as the α -gal A activity was extremely low and lyso-Gb₃ concentration was 11.9 ng/ml. The patient is suffering of LVH, increased pulmonary artery diameter, dilatation of the ascending aorta and aortic valve stenosis, because of which he underwent a valve replacement surgery. Ophthalmological evaluation indicated lipid deposition with blurriness of the cornea. The patient is currently under ERT. His 34-year-old brother was identified with the same mutation and has extremely low α -gal A activity, lyso-Gb3 concentration 12.9 ng/ml and albuminuria of no other aetiology.

Lastly the c.453C>G mutation (Y151X - p.Tyr151Ter) in exon 3 of the GLA gene was revealed in three members (55 \pm 16, range 42–73 years) of an other Greek family. The proband, a 50-year-old male, was initially misdiagnosed at the age of 36 with SLE, as he suffered from small joints arthralgia both in hands and feet, low-grade fever and positive antinuclear antibodies (ANA). A kidney biopsy performed at the time due to proteinuria (2.4 gr/24h) and deteriorated kidney function, indicated a possible SLE nephritis (class III – WHO). He submitted on treatment for SLE for some years without clinical benefit. However angiokeratomas in the pelvic area with which the patient was presented were progressing over the time. This finding necessitated the revaluation of the patient. Kidney biopsy showed cytoplasmic microvacuolization of enlarged podocytes, as well as segmental sclerosis in some glomeruli. The α-gal A activity was found pathological low (0.06 μmol/l/h). SLE treatment stopped and the patient commenced ERT. At that time the patient presented pain crisis, acroparesthesias, hypohidrosis, temperature intolerance, skin lesions (angiokeratomas), cornea verticillata corneopathy, nephropathy and mitral valve prolapse / insufficiency. The proband's mother (73 years old) presented at the age of 64 with multiple parapelvic kidney cysts, nephropathy, mitral valve prolapse, rhythm abnormalities, LVH and WML on brain MRI. She was also suffering of chronic chough and depression. The probands sister was misdiagnosed too, as years ago she was reporting severe joint pains attributed to mixed connective tissue disease (MCTD). After the probands diagnosis and at the age of 41 she genotyped and, at that time, the GLA mutation was found. Thereafter, she presented slight albuminuria, mitral valve prolapse, mitral and aortic valve insufficiency. Autonomic and central nervous system where also affected, as the patient reported acroparesthesias, temperature intolerance and tinnitus. All three patients were diagnosed with cornea verticillata corneopathy and are currently under ERT.

DISCUSSION

The D313Y mutation

Contradicting results about the pathogenicity of this mutation have been reported in the literature since its first description on 1993 [14]. The mutation has been detected in many series of patients presenting signs of FD [15, 16, 17, 18, 19, 20, 21, 22]. However, Niemann et al. [23] describes this variant as non pathogenic, although his two patients were presenting decreased α -gal A activity. Similarly, Oder et al. [24] supports that the D313Y genotype does not lead to severe organ manifestations as seen in genotypes known to be causal for classical FD and Froissart et al. [25] characterizes the mutation as "pseudodeficient allele" implying that it is a sequence variant which encodes an enzyme that is transported to the lysosomes, where it has about 75% of normal enzymatic activity. The D313Y mutation has been also referred as polymorphism [17], despite that, according to the ExAC and 1000 Genomes databases, its frequency in the World and European population is below 1%. Finally, it must be mentioned that by bioinformatics analysis this mutation is predicted as probably damaging (PolyPhen-2) or damaging (SIFT).

In our study five of seventeen carriers of the D313Y mutation (54 \pm 9, range 45–65 years) proved as suffering definite FD according to the recently published criteria of the disease [13]. The presentation of the disease in our patients indicates that the mutation results in a milder phenotype, with later onset of symptoms. This phenotype, also including milder mono- or oligosymptomatic cases [17], is characterised as atypical or type 2 [26]. The late onset of clinical symptoms and the milder than the typical phenotype of FD in these patients can be partly explained by the high α -gal A residual activity, since there is evidence that the mutated α -gal A reaches intracellularly the lysosomes [27]. Actually, in nearly all male D313Y carriers of our study (61 \pm 11, range 45–78 years) α -gal A activity was decreased in a range of 56.2-87.5% comparing to normal. Another reason for the decreased activity of the D313Y enzyme in plasma could be a functional intolerance to blood plasma neutral pH

conditions. This effect is irreversible and, once in contact with a neutral or basic pH environment D313Y enzyme remains inactive, even if transferred to optimal pH [19, 27].

Interestingly, the vast majority of our D313Y patients were presenting with neurological symptoms and signs. Moreover, one of them was misdiagnosed as MS while two carriers of the mutation, in whom definite FD diagnosis has not been established as yet, are diagnosed and treated as MS. Similarly the detailed study of the renal biopsy in one of our cases underlies the significance of early detection of Fabry specific findings in cases with FSGS.

Novel mutations

The pathogenicity of the above mentioned novel mutations has been undoubtfully proved as all carriers suffer definite FD. Of them the c.835C>T (p.Gln279Ter, Q279X) is a nonsense mutation causing an interruption of the reading frame by a premature stop codon, which results in a truncated protein. A truncated protein and the subsequent loss of its functionality is strong evidence that the mutation is probably pathogenic for FD [12]. No amino acid change at this position has ever been described in NCBI and Fabry Database (http://fabry-database.org/) [28]. Accordingly the c.453C>G (p.Tyr151Ter, Y151X) mutation causes an interruption of the reading frame by a premature stop codon, which results in a truncated protein and subsequent loss of its functionality. This is a strong evidence that the mutation could be pathogenic for FD [12]. A further evidence that this mutation is pathogenic is provided by the fact that the same amino acid change at this position but in a different nucleotide has already been described as disease-causing [29, 17].

As far as the c.280T>A (C94S), c.924A>C (K308N), c.511G>A (G171S) mutations are considered, bioinformatics analysis supports their disease causing effect as they are predicted as probable damaging (PolyPhen-2) and/or damaging (SIFT). A further evidence of their pathogenicity could be the fact that the same aminoacid changes at the corresponding

positions but in different nucleotides have already been described as disease causing [30, 31, 29,].

CONCLUSIONS

We report five novel *GLA* mutations causing FD. Moreover, we offer strong evidence that the D313Y mutation could be pathogenic. It seems that this mutation is related with a later-onset milder than the typical phenotype with normal lysoGb₃ concentration.

Additionally our study underlines the significance of family members genotyping, newborn screening and genetic counselling in avoiding misdiagnoses and crucial delays of diagnosis and treatment of the disease. Finally we confirmed the fact that heterozygous females may develop mild to severe FD as well as that genotype-phenotype correlation does not exist even among the members of the same families.

Table 1. Characteristics of patients carrying the D313Y mutation.

Patient / Gender / Age (years)	FD related clinical findings	Previous diagnosis
1 / M / 52	End stage renal disease – dialysis, acroparesthesias, renal	FSGS
1,111,02	cysts, elevated plasma and urine Gb ₃ concentration	nephropathy
2/F/30	Healthy	no
3/F/70	Healthy	no
4/F/46	Three strokes of unknown origin, WML on brain MRI,	no
	micro-albuminuria, oedema, acroparesthesias, elevated urine Gb ₃ concentration	
5/F/65	Multiple WML on brain MRI, depression, elevated plasma	MS, RA,
	Gb ₃ concentration	spastic
		quadriplegia
6/F/62	Acroparesthesias, GI symptoms since adolescence	no
7 / M / 45	End stage renal disease on dialysis, WML and vertebrobasilar	Nephropathy
	vessel changes on brain MRI, increased cardiac	of unknown
	interventricular septum echogenicity, hearing loss of higher	origin
	frequencies	
8 / M / 62	End stage renal disease on dialysis	no
9/F/36	WML on brain MRI, acroparesthesias	MS
10 / M / 78	End stage renal disease on dialysis	RA
11 / F / 47	LVH, CKD	no
12 / M / 68	No FD signs	Myopathy
13 / F / 46	No FD signs	NMO
14 / F / 27	WML on brain MRI	MS
15 / M / 61	End stage renal disease on dialysis	Diabetic
		nephropathy
16 / M / 63	CKD, hearing loss, multiple ischaemic infracts on brain MRI	no
17 / F / 59	Cornea verticillata, hearing loss, LVH, acroparesthesias, GI	no
	symptoms (pain – diarrhoea) since adolescence, hypohidrosis,	
	T2 -WML or ischaemic infracts on brain MRI	

Table 2. Clinical characteristics of patients with the five novel mutations.

Mutation	Clinical data	
(NM_000169.2)		
c.835C>T	CKD, haematuria, proteinuria, pain in extremities, dyshidrosis,	
p.Gln279Ter	angiokeratomas, zero activity of a-gal A, LVH, kidney biopsy consistent	
Q279X	with FD, TIA, pathological elevated lyso-Gb ₃	
c.280T>A	Microalbuminuria, end stage renal disease – dialysis, cornea verticillata	
p.Cys94Ser	corneopathy, acroparesthesias, dyshidrosis, LVH, WML on brain MRI,	
C94S	zero activity of a-gal A, pathological elevated lyso-Gb ₃ , pathological	
	elevated Gb ₃ in plasma/urine	
c.924A>C	CKD, haematuria, kidney biopsy findings related to FD, zero activity of	
p.Lys308Asn	a-gal A, LVH	
K308N		
c.511G>A	Proteinuria, end stage renal disease – dialysis, LVH, valvulopathy, cornea	
p.Gly171Ser	verticillata corneopathy, extremely low activity of a-gal A, pathological	
G171S	elevated lyso-Gb ₃	
c.453C>G	Proteinuria, end stage renal disease – dialysis, kidney biopsy findings	
p.Tyr151Ter	related to FD, pain in extremities – acroparesthesias, severe	
Y151X	angiokeratomas, hypohidrosis – temperature intolerance, cornea	
	verticillata corneopathy, LVH, valvulopathy, rhythm abnormalities,	
	WML on brain MRI, tinnitus, low activity of a-gal A	

D313Y mutation of the <i>GLA</i> . (a) Glomerulus with segmental sclerosis - PAS X 4 Segmental sclerosis with features of the "collapsing" variant - Jones' silver X 40 appearing glomerulus with a small area of sclerosis adhering to Bowman's capsu 400. (d) Cytoplasmic microvacuolization of podocytes, suggestive of FD - PAS Figure 2. Sanger confirmation of novel mutations Figure 3. Electron microscopy findings of renal biopsy from a male FD patient Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding marked with black arrows in (a) methylene blue semithin section, (b) tubulation fibroblast and (c,d) podocytes.	arrying the
appearing glomerulus with a small area of sclerosis adhering to Bowman's capsu 400. (d) Cytoplasmic microvacuolization of podocytes, suggestive of FD - PAS Figure 2. Sanger confirmation of novel mutations Figure 3. Electron microscopy findings of renal biopsy from a male FD patien Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical findinarked with black arrows in (a) methylene blue semithin section, (b) tubula fibroblast and (c,d) podocytes.	400. (b)
400. (d) Cytoplasmic microvacuolization of podocytes, suggestive of FD - PAS Figure 2. Sanger confirmation of novel mutations Figure 3. Electron microscopy findings of renal biopsy from a male FD patien Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical findi marked with black arrows in (a) methylene blue semithin section, (b) tubula fibroblast and (c,d) podocytes.	00. (c) Pale
Figure 2. Sanger confirmation of novel mutations Figure 3. Electron microscopy findings of renal biopsy from a male FD patien Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical findi marked with black arrows in (a) methylene blue semithin section, (b) tubula fibroblast and (c,d) podocytes.	ule - PAS X
Figure 3. Electron microscopy findings of renal biopsy from a male FD patien Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding marked with black arrows in (a) methylene blue semithin section, (b) tubulation fibroblast and (c,d) podocytes.	S X 400.
Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding marked with black arrows in (a) methylene blue semithin section, (b) tubulation fibroblast and (c,d) podocytes.	
marked with black arrows in (a) methylene blue semithin section, (b) tubula fibroblast and (c,d) podocytes.	nt carrying the
fibroblast and (c,d) podocytes.	ing of FD, are
	ar cells and a

322	Abbreviations
323	$\alpha\text{-gal}$ A: Enzyme $\alpha\text{-galactosidase}$ A; ACMG: American College of Medical Genetics and
324	Genomics; ANA: Antinuclear antibodies; CKD: Chronic kidney disease; DBS: Dried blood
325	spot; ERT: Enzyme replacement therapy; FD: Fabry disease; FSGS: focal and segmental
326	glomerulosclerosis; GI: Gastrointestinal; GLA: α -galactosidase A gene; GL-3 or Gb3:
327	globotriaosylceramide; LVH: Left ventricular hypertrophy; lyso-Gb3:
328	globotriaosylsphingosine; MRI: Magnetic resonance imaging; MV: mean value; MS: Multiple
329	sclerosis; NMO: Neuromyelitis optica; RA: Rheumatoid arthritis; SD: Standard deviation;
330	SLE: Systemic lupus erythematosus; TIA: Transient ischemic attack; UT: Under treatment;
331	VUS: Variant of uncertain significance; WML: White matter lesions;
332	Acknowledgements
333	We appreciate the help and advice from A. Andrikos, S. Kouzouka, A. Triantafillou, D.
334	Kaltsidou, A. Kaliantzoglou, A. Lysitska, K. Papadopoulos, K. Zoganas, G. Tsivgoulis, Z.
335	Tegou, A. Fountoglou, T. Karapanagiotidis, S. Patsialas.
336	Availability of data and materials
337	All data supporting our findings are included in the manuscript.
338	Authors' contributions
339	KK: collection, analysis and interpretation of the data, pedigree analysis, literature review,
340	drafting and revision of the manuscript. KS: Renal biopsy and analysis on electron
341	microscope. PP: Renal biopsy and analysis on optical microscope. MS, MZ and GL:
342	genotyping and bioinformatics analysis. KS, PP, EM, PK, CK, AO, JK: treating physicians of
343	patients. AEG: design and coordination of the study, revision of the manuscript. All authors
344	approved the final version of the manuscript.

345	Competing interests
346	KK received travel assistance from Shire and Genzyme and speaker's honoraria from Shire.
347	KS received travel assistance and speaker's honoraria from Shire. PP and AO received travel
348	assistance from Genzyme and Shire. CK received travel assistance from Shire. AEG received
349	research grants from Shire. The authors MZ, GL, MS, EM, PK, JK, report no competing
350	interests.
351	Funding
352	The study has been partially supported by a grant from the Research Committee of the
353	University of Thessaly.
354	Consent for publication
355	Written informed consents for this publication were obtained from patients. A copy of the
356	consent form is available for review by the Editor of this journal.
357	Ethics approval and consent to participate

- This report was approved by the institutional review board of the University of Thessaly,
- Larissa. Written informed consents for participation were obtained from patients.

361 References

- 1. Germain DP. Fabry disease. Orphanet J Rare Dis. 2010;5:30
- 2. Schiffmann R. Fabry disease. Pharmacol Ther. 2009;122:65-77
- 1000 Genomes Project Consortium, Abecasis GR, Auton A, Brooks LD, DePristo MA,
 Durbin RM, et al. An integrated map of genetic variation from 1,092 human genomes.
 Nature. 2012;491:56–65
- Barba-Romero M-A, Rivera-Gallego A, Pintos-Morell: Spanish FOS-Study Group. Int J Clin Pract. 2011;65:903–10
- 5. Laney DA, Fernhoff PM. Diagnosis of Fabry disease via analysis of family history. J Genet Couns. 2008;17:79–83
- Mehta A, Ricci R, Widmer U, Dehout F, Garcia de Lorenzo A, Kampmann C, et al.
 Fabry disease defined: baseline clinical manifestations of 366 patients in the Fabry
 Outcome Survey. Eur J Clin Invest. 2004;34:236–42
- Gal A, Hughes DA, Winchester B. Towards a consensus in the laboratory diagnostics of Fabry disease – recommendations of a European expert group. J Inherit Metab Dis. 2011;34:509–14
- 8. Winchester B, Young E. Biochemical and genetic diagnosis of Fabry disease. In: Mehta A, Beck M, Sunder-Plassmann G, editors. Fabry disease: perspectives from 5 years of FOS. Oxford: Oxford PharmaGenesis; 2006. Chapter 18.
- Hughes DA, Evans S, Milligan A, Richfield L, Mehta A. A multidisciplinary approach to the care of patients with Fabry disease. In: Mehta A, Beck M, Sunder-Plassmann G, editors. Fabry disease: Perspectives from 5 years of FOS. Oxford: Oxford PharmaGenesis; 2006. Chapter 35
- Laney DA, Fernhoff PM. Diagnosis of Fabry disease via analysis of family history. J Genet Couns. 2008;17:79–83

- 11. Aerts JM, Groener JE, Kuiper S, Donker-Koopman WE, Strijland A, Ottenhoff R, et al. Elevated globotriaosylsphingosine is a hallmark of Fabry disease. Proc Natl Acad Sci USA. 2008;105:2812–7
- 12. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17:405-24
- 13. Biegstraaten M, Arngrímsson R, Barbey F, Boks L, Cecchi F, Deegan P, et al. Recommendations for initiation and cessation of enzyme replacement therapy in patients with Fabry disease: the European Fabry Working Group consensus document. Orphanet J Rare Dis. 2015; doi:10.1186/s13023-015-0253-6
- 14. Eng CM, Resnick-Silverman LA, Niehaus DJ, Astrin KH, Desnick RJ. Nature and frequency of mutations in the alpha-galactosidase A gene that cause Fabry disease. Am J Hum Genet. 1993;53:1186-97
- 15. Gaspar 2010 Gaspar P, Herrera J, Rodrigues D, Cerezo S, Delgado R, Andrade CF, et al. Frequency of Fabry disease in male and female haemodialysis patients in Spain. BMC Med Genet.2010; doi: 10.1186/1471-2350-11-19
- 16. Wang RY, Bodamer OA, Watson MS, Wilcox WR; ACMG Work Group on Diagnostic Confirmation of Lysosomal Storage Diseases. Lysosomal storage diseases: diagnostic confirmation and management of presymptomatic individuals. Genet Med. 2011; doi: 10.1097/GIM.0b013e318211a7e1
- 17. Lukas J, Giese AK, Markoff A, Grittner U, Kolodny E, Mascher H, et al. Functional Characterisation of alpha-galactosidase a mutations as a basis for a new classification system in fabry disease. PLoS Genet. 2013; doi:10.1371/journal.pgen.1003632

- Böttcher T, Rolfs A, Tanislav C, Bitsch A, Köhler W, Gaedeke J, et al. Fabry disease underestimated in the differential diagnosis of multiple sclerosis? PLoS One.2013; doi: 10.1371/journal.pone.0071894
- 19. Lenders M, Duning T, Schelleckes M, Schmitz B, Stander S, Rolfs A, et al. Multifocal white matter lesions associated with the D313Y mutation of the a-galactosidase A gene. PLoS One. 2013; doi: 10.1371/journal.pone.0055565
- Samuelsson K, Kostulas K, Vrethem M, Rolfs A, Press R. Idiopathic small fiber neuropathy: phenotype, etiologies and the search for fabry Disease. J Clin Neurol. 2014;10(2):108-118
- Becker J, Rolfs A, Karabul N, Berlit P, Kraemer M. D313Y mutation in the differential diagnosis of white matter lesions: Experiences from a multiple sclerosis outpatient clinic. Mult Scler. 2016;22:1502-5
- 22. De Brabander I, Yperzeele L, Groote CC, Brouns R, Baker R, Belachew S, et al. Phenotypical Characterization of α -Galactosidase a Gene Mutations Identified in a Large Fabry Disease Screening Program in Stroke in the Young. Clin Neurol Neurosurg. 2012;115:1088-93
- 23. Niemann M, Rolfs A, Giese A, Mascher H, Breunig F., Ertl G, et al. Lyso-Gb3 indicates that the Alpha-Galactosidase A mutation D313Y is not clinically relevant for Fabry Disease. JIMD Rep. 2012; doi: 10.1007/8904_2012_154
- 24. Oder D, Üceyler N, Liu D, Hu K, Petritsch B, Sommer C, et al. Organ manifestations and long-term outcome of Fabry disease in patients with the GLA haplotype D313Y. BMJ Open. 2016; doi: 10.1136/bmjopen-2015-010422
- 25. Froissart R, Guffon N, Vanier MT, Desnick RJ, Maire I. Fabry disease: D313Y is an alpha-galactosidase A sequence variant that causes pseudodeficient activity in plasma. Mol Genet Metab. 2003;80:307–14

- 26. Lenders M, Weidemann F, Kurschat C, Canaan-Kühl S, Duning T, Stypmann J, et al.

 Alpha-Galactosidase A p.A143T, a non-Fabry disease-causing variant. Orphanet J

 Rare Dis. 2016;11:54
- 27. Yasuda M, Shabbeer J, Benson SD, Maire I, Burnett RM, Desnick RJ. Fabry disease: characterization of alpha-galactosidase A double mutations and the D313Y plasma enzyme pseudodeficiency allele. Hum Mutat. 2003;22:486–92
- 28. Meiji Pharmaceutical university. http://fabry-database.org. Accessed 15 February 2017
- 29. Shabbeer J, Yasuda M, Benson SD, Desnick RJ. Fabry disease: Identification of 50 novel α-galactosidase mutations causing the classic phenotype and three-dimensional structural analysis of 29 missense mutations. Hum Genomics. 2006;2:297-309
- 30. Blaydon D, Hill J, Winchester B. Fabry disease: 20 novel GLA mutations in 35 families. Hum Mutat. 2001;18:459
- 31. Shimotori M, Maruyama H, Nakamura G, Suyama T, Sakamoto F, Itoh M, et al. Novel mutations of the GLA gene in Japanese patients with Fabry disease and their functional characterization by active site specific chaperone. Hum Mutat. 2008;29:331

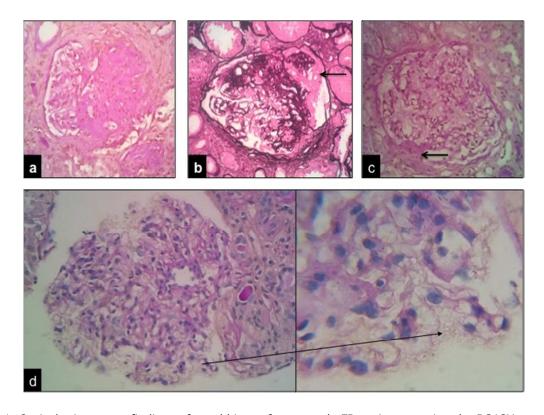


Figure 1. Optical microscopy findings of renal biopsy from a male FD patient carrying the D313Y mutation of the GLA. (a) Glomerulus with segmental sclerosis - PAS X 400. (b) Segmental sclerosis with features of the "collapsing" variant - Jones' silver X 400. (c) Pale appearing glomerulus with a small area of sclerosis adhering to Bowman's capsule - PAS X 400. (d) Cytoplasmic microvacuolization of podocytes, suggestive of FD - PAS X 400.

224x167mm (72 x 72 DPI)

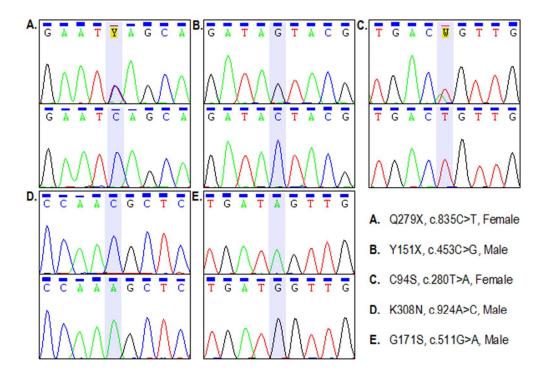


Figure 2. Sanger confirmation of novel mutations.

158x112mm (96 x 96 DPI)

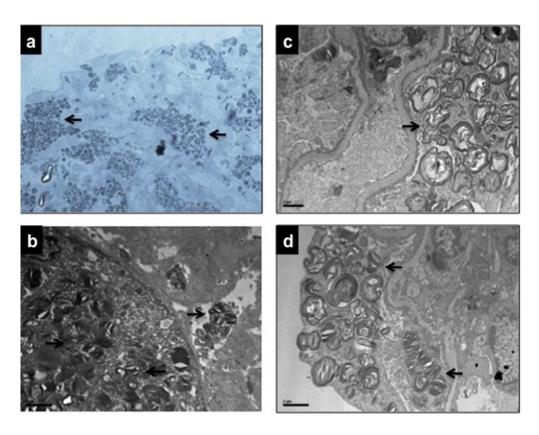


Figure 3. Electron microscopy findings of renal biopsy from a male FD patient carrying the Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding of FD, are marked with black arrows in (a) methylene blue semithin section, (b) tubular cells and a fibroblast and (c,d) podocytes.

178x139mm (72 x 72 DPI)

Koulousios et al. STROBE Research Checklist

	Item No	
Title and abstract	1	Title [Page 1]: Fabry disease due to D313Y and novel GLA mutations
		Abstract [Page 3]
		The study is a polycentric observational cohort study which addresses the question of
		pathogenicity of specific Fabry disease mutations, using cohort data.
		Abstract [Page 3]: The abstract summarises the objectives, settings and participants
		primary and secondary outcome measures, results and conclusions of the study.
Introduction		
Background/rationale	2	Introduction [Page 4]
		Introduction to the scientific background of Fabry disease and <i>GLA</i> mutations.
Objectives	3	Introduction [Page 5]
		Aim of the study is to report five novel <i>GLA</i> mutations resulting in FD and provide
		evidence of pathogenicity of the D313Y mutation regarding which contradictory data
		have been presented in the literature.
Methods		
Study design	4	Methods - Study population [Page 5]
		- Polycentric observational cohort study, multiple patient cases from various medical
		units all over Greece (polycentric study).
		- The study was approved by the institutional review board of the University of
		Thessaly, Larissa.
Setting	5	Methods - Study population and Clinical assessment [Page 5]
		- Patient cases from multiple medical units in various locations all over Greece.
		- Family members of the majority of the Greek FD patients, who were diagnosed
		during the last years.
		- Pedigree analysis after personal interviews with the selected individuals in their
		places of residence.
Participants	6	Methods - Study population and Clinical assessment [Page 5]
		Participants of the study fulfilled one of the following eligibility criteria:
		i) relatives of a patient with definite FD diagnosis.
		ii) patient cases with nephrological, cardiac or neurological symptoms suspicious of
		FD diagnosis from various medical units all over Greece.
		- Additional genotyping of 145 apparently healthy subjects.
		- The source of participants selection was their medical records (relation with a FD
		patient, patients with FD related symptoms), analysed in collaboration with their
		treating physicians.
		- Patients follow-up in collaboration with their treating physicians. Revaluation of the
		clinical status of each selected individual.
Variables	7	Clinical assessment - Laboratory evaluation - Genotyping – [Pages 5-6]

Variables used in the study:

- i) Heart or kidney disease, cerebrovascular events, death at young age and respective causes of death.
- ii) FD related cardiac, renal and neurological signs and symptoms.
- iii) Familiar relationship to a FD patient.
- iv) Enzyme α-Gal A activity in all male subjects.
- v) Lyso-Gb₃ in all subjects.
- vi) Plasma and urine Gb₃ concentration in selected cases.
- vii) Presence of a GLA mutation.
- viii) Verification of novel mutations.
- ix) Evaluation of kidney biopsies.

Diagnostic criteria [Page 6]

The diagnosis of FD was posed according to the recently published criteria of the disease [Biegstraaten et al. 2015]

Data sources/ measurement

8 Clinical assessment - Laboratory evaluation - Genotyping - page 5-6

Method of assessment for each variable:

- i) Medical history.
- ii) Physical examination.
- iii) Pedigree analysis.
- iv) In DBS by tandem mass spectrometry.
- v) In DBS by HPLC and tandem spectrometry.
- vi) Tandem mass spectrometry.
- vii) Genotyping.
- viii) Standard Sanger sequencing using Variant Reporter software v1.1 (Applied Biosystems).
- ix) Optical and electron microscopy.

Bias

9 Study population [Page 5]

Cross examination for the presence of D313Y mutation by genotyping of 145 apparently healthy subjects.

Laboratory evaluation [Page 5]

Measurement of α -Gal A activity only in male subjects.

Genotyping [Page 6]

Exclusion of common polymorphisms (UCSC Common SNPs) by bioinformatic analysis using PhyloP, SIFT, Grantham and PolyPhen tools, in comparison to their global (1000 Genomes Global Minor Allele Frequency, ExAC) and European frequency (5000 Exomes European Minor Allele Frequency).

Study size

10 Study population [Page 5]

The cohort size of 62 subjects was arrived after selection of interesting patient cases between families of 9 unrelated patients with definite diagnosis of FD as well as amongst cases with nephrological, cardiac or neurological symptoms suspicious of this diagnosis. Eighteen family members of the last cases were also examined after the confirmation of diagnosis.

Quantitative variables

Laboratory evaluation [Page 5]

In DBS, α -Gal A activity was measured in all male subjects by tandem mass spectrometry, lyso-Gb₃ in all subjects by HPLC and tandem spectrometry as well as plasma and urine Gb₃ concentration in selected cases by tandem mass spectrometry.

Statistical methods

Age of each patient or patient group is presented either as an absolute number or as mean value \pm the Standard Deviation (mean \pm SD).

Results

Participants

13 Results [Page 6]

62 subjects were potentially eligible for participation in the study and were genotyped for a *GLA* mutation, clinically analysed and under follow-up for a significant period of time.

Abstract [page 3]

The 62 subjects were selected between 3 categories of patient cases:

BMJ Open

- i) Family members of unrelated patients with definite FD diagnosis.
- ii) Clinically suspected cases.
- iii) Family members of the previous category.

Descriptive data

14 Abstract [Page 3]

- i) 25 family members of 9 unrelated patients with definite FD diagnosis.
- ii) Ten clinically suspected cases (nephrological, cardiac or neurological symptoms suspicious of FD diagnosis) from various medical units all over Greece.
- iii) 18 family members of the previous category.

Results [Pages 6-11]

- i) Patients carrying the D313Y mutation: 17 (54±14, range 27–78 years). [page 6]
- ii) Male patients carrying the D313Y mutation: 7 (61±11, range 45–78 years). [page 6]
- iii) Female patients carrying the D313Y mutation: 10 (49±15, range 27–70 years). [page 6]
- iv) Patients carrying novel *GLA* mutations: 16 members (42±19, range 1–73 years) of 5 unrelated families, all fulfilling the diagnostic criteria of a definite diagnosis of FD. [page 8]
- v) Patients identified with the novel c.835C>T mutation: 4 members (30±24, range 1–60 years) of a Greek family. Proband of the family: a 31-year-old male with typical FD clinical and laboratory phenotype. [page 8,9]
- vi) Patients identified with the novel c.280T>A mutation: 4 members (46±11, range 30–56 years) belonging to another Greek family. Proband of the family: a 48-year-old female with typical FD clinical and laboratory phenotype. [page 9]
- vii) Patients identified with the novel c.924A>C mutation: 3 members (44±26, range 17–69 years) of a Greek family. Proband of the family: a 46-year-old male with typical FD clinical and laboratory phenotype. [page 9,10]
- viii) Patients identified with the novel c.511G>A mutation: 2 members $(37 \pm 4, \text{ range } 34-39 \text{ years})$ of an Albanian family living in Greece. Proband of the family: a 39-year-old male with typical FD clinical and laboratory phenotype. **[page 10]**
- ix) Patients identified with the novel c.453C>G mutation: 3 members (55 ± 16 , range 42–73 years) of an other Greek family. Proband of the family: a 50-year-old male with typical FD clinical and laboratory phenotype. [page 10,11]

Outcome data

5 Results [pages 6-11]

i) Six (all with definite FD) out of the 62 genotyped subjects were carrying four previously described *GLA* mutations. **[page 6]**

ii) The D313Y mutation was revealed in 17 individuals, 7 males and 10 females. **[page 6]** iii) The novel c.835C>T, c.280T>A, c.924A>C, c.511G>A and c.453C>G *GLA* mutations were detected in 16 members of 5 unrelated families.

Main results 16 **Results [pages 6-11]**

- i) Six (all with definite FD) out of the 62 genotyped subjects were carrying four previously described *GLA* mutations. **[page 6]**
- ii) The D313Y mutation was revealed in 17 individuals, 7 males and 10 females. [page 6]
- iii) The D313Y mutation was revealed in none of the healthy subjects. [page 6]
- iv) The diagnosis of FD was definitely posed in 5 carriers of the D313Y mutation. [page 6-8]
- v) Two D313Y mutation carriers were patients presenting no FD related signs or symptoms. [page 8]
- vi) Eight D313Y mutation carriers were patients presenting other FD signs mainly neurological that, however, can not document a definite diagnosis of FD. [page 8]
- vii) Two D313Y mutation carriers were apparently healthy. [page 8]
- viii) The diagnosis of FD was definitely posed in all carriers of the novel mutations reported.

[page 8-11]

Other analyses

- i) Bioinformatics analysis of every GLA mutation reported, by (PolyPhen-2) or (SIFT) systems. [page 12-13]
 - ii) Novel mutations verification by standard Sanger sequencing using Variant Reporter software v1.1 (Applied Biosystems). [page 6]

Discussion

Key results

- 18 i) Five novel *GLA* mutations causing classical Fabry Disease are reported. [page 14]
 - ii) Strong evidence that the D313Y mutation could be pathogenic is offered. [page 14]

Limitations

19 The main limitation is the lack of detailed clinical data in older participants. [page 4]

Interpretation

20 Discussion [Pages 11-13]

- Five of seventeen carriers of the D313Y mutation proved as suffering definite FD according to the recently published criteria of the disease. The presentation of the disease in our patients indicates that the mutation results in a milder phenotype, with later onset of symptoms. [page 12]
- Contradicting results about the pathogenicity of this mutation have been reported in the literature since its first description on 1993. [page 11-12]
- The mutation has been detected in many series of patients presenting signs of FD. However, Niemann et al. describes this variant as non pathogenic, although his two patients were presenting decreased α-gal A activity. Similarly, Oder et al. supports that the D313Y genotype does not lead to severe organ manifestations as seen in genotypes known to be causal for classical FD and Froissart et al. [i] characterizes the mutation as "pseudodeficient allele" implying that it is a sequence variant which encodes an enzyme that is transported to the lysosomes, where it has about 75% of normal enzymatic activity. The D313Y mutation has been also referred as polymorphism. [page 11-12]
- The vast majority of the D313Y patients of the study were presenting with neurological symptoms and signs. [page 12]
- Kidney biopsy is significant for a FD diagnosis in patients with renal manifestations of the disease [page 7, 9-11]

- The pathogenicity of the five novel mutations reported has been undoubtfully proved as all carriers suffer definite FD. [page 13]

Generalisability 21 **Discussion [Pages 11-13] Conclusion [Page 13-14]**

The study underlines the significance of:

- i) Diagnosis of FD and possible treatment of patients carrying the 5 novel and the D313Y *GLA* mutations.
- ii) Family members genotyping.
- iii) Newborn screening and genetic counselling.
- iv) Avoiding misdiagnoses and crucial delays of diagnosis and treatment of the disease.

The study confirmed the facts that:

- i) Heterozygous females may develop mild to severe FD.
- ii) Misdiagnosis is common in FD.
- iii) Genotype-phenotype correlation does not exist even among the members of the same families.

Other information

Funding

22 Funding [Page 19]

The study has been partially supported by a grant from the Research Committee of the University of Thessaly.

BMJ Open

Fabry disease due to D313Y and novel GLA mutations

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-017098.R1
Article Type:	Research
Date Submitted by the Author:	14-Jun-2017
Complete List of Authors:	Koulousios, Konstantinos; University of Thessaly Faculty of Medicine, Stylianou, Konstantinos; University Hospital of Heraklion Pateinakis, Panagiotis; Geniko Nosokomeio Thessalonikis Papageorgiou Zamanakou, Maria; CeMIA SA Loules, Gedeon; CeMIA SA Manou, Eleni; Geniko Nosokomeio Thessalonikis Papageorgiou Kyriklidou , Parthena; Geniko Nosokomeio Thessalonikis Papageorgiou Katsinas, Christos; Mpodosakeio General Hospital Ouzouni, Alexandra; Geniko Nosokomeio Kabalas Kyriazis, John; Geniko Nosokomeio Chiou Skylitseio Speletas , Matthaios ; University of Thessaly Faculty of Medicine Germenis , Anastasios E. ; University of Thessaly Faculty of Medicine
Primary Subject Heading :	Genetics and genomics
Secondary Subject Heading:	Renal medicine, Cardiovascular medicine, Neurology, Diagnostics
Keywords:	Fabry disease, D313Y GLA mutation, Novel GLA mutations, Kidney biopsy, Misdiagnosis

SCHOLARONE™ Manuscripts

1 Fabry disease due to D313Y and novel GLA mutations

- 2 Konstantinos Koulousios^{a*}, Konstantinos Stylianou^b, Panagiotis Pateinakis^c, Maria
- 3 Zamanakou^d, Gedeon Loules^d, Eleni Manou^c, Parthena Kyriklidou^c, Christos Katsinas^e,
- 4 Alexandra Ouzouni^f, John Kyriazis^g, Matthaios Speletas^a, Anastasios E. Germenis^a
- 5 *Corresponding author
- 6 ^a Department of Immunology & Histocompatibility, School of Medicine, University of
- 7 Thessaly, Panepistimiou 3, GR-41500 Biopolis, Larissa, Greece
- 8 b University Hospital of Heraklion, Heraklion, Greece
- 9 ^c Papageorgiou General Hospital, Thessaloniki, Greece
- 10 d CeMIA SA, Makriyianni 31, GR-41334 Larissa, Greece
- 11 ^e Mpodosakeio General Hospital, Ptolemaida, Greece
- 12 ^f Kavala General Hospital, Kavala, Greece
- 13 ^g Chios General Hospital, Chios, Greece
- 14 Konstantinos Koulousios (konkoul@gmail.com)
- 15 Konstantinos Stylianou (kstylianu@gmail.com)
- Panagiotis Pateinakis (pateinakis@hotmail.com)
- 17 Maria Zamanakou (mzamanakou@cemia.eu)
- 18 Gedeon Loules (gedloules@cemia.eu)
- 19 Eleni Manou (elmanou@webcoreforscience.com)
- 20 Parthena Kyriklidou (renakiri@yahoo.it)
- 21 Christos Katsinas (katsinas@otenet.gr)
- 22 Alexandra Ouzouni (alouzouni2808@gmail.com)
- John Kyriazis (jks@otenet.gr)
- Matthaios Speletas (maspel@med.uth.gr)

Anastasios E. Germenis (agermen@med.uth.gr)



27	Abstract
28	Objectives: Our aim is to report four novel <i>GLA</i> mutations resulting in FD and provide
29	evidence of pathogenicity of the D313Y mutation regarding which contradictory data have
30	been presented in the literature.
31	Setting and participants: 25 family members of nine unrelated patients with definite FD
32	diagnosis, ten clinically suspected cases and eighteen members of their families were included
33	in this polycentric cohort study.
34	Primary and secondary outcome measures: Genotyping and measurement of lyso-Gb ₃ was
35	performed in all individuals. The α -Gal A activity was measured in all males as well as
36	plasma and urine Gb ₃ concentration in selected cases. Optical and electron microscopy was
37	performed in kidney biopsies of selected patients. All the above were evaluated in parallel
38	with the clinical data of the patients.
39	Results: Fourteen new cases of FD were recognised, four of which were carrying already
40	described GLA mutations. Four novel GLA mutations, namely c.835C>T, c.280T>A,
41	c.924A>C, and c.511G>A, resulting in a classical FD phenotype were identified. Moreover,
42	FD was definitely diagnosed in five patients carrying the D313Y mutation. Eight D313Y
43	carriers were presenting signs of FD despite not fulfilling the criteria of the disease, two had
44	no FD signs and two others were apparently healthy.
45	Conclusions: Four novel GLA pathogenic mutations are reported and evidence of
46	pathogenicity of the D313Y mutation is provided. It seems that the D313Y mutation is
47	related with a later-onset milder than the typical phenotype with normal lysoGb ₃
48	concentration. Our study underlines the significance of family members genotyping and
49	newborn screening in avoiding misdiagnoses and crucial delays of diagnosis and treatment of
50	the disease.

Strengths and limitations of this study

- This is the largest series in the literature of clinically evaluated male and female carriers of the *GLA* D313Y mutation supporting its possible pathogenicity that occasionally has been proved by renal biopsy.
- Novel GLA mutations resulting to a classical Fabry disease phenotype are presented.
- The main limitation is the lack of detailed clinical data in older participants.
- Biopsies of affected organs, the gold standard of definite diagnosis, are not available in all cases.

INTRODUCTION

FD or Anderson-Fabry disease is an X-linked inherited metabolic disorder, that results from mutations in the α -gal A gene (*GLA* gene), leading to reduction of the enzyme activity and subsequent accumulation of Gb₃ (or GL-3) in plasma, urine and cellular lysosomes throughout the body. These depositions cause a multisystemic pathology with life-threatening manifestations, including renal failure, cardiac and cerebrovascular disease [1, 2].

More than 900 currently known *GLA* mutations have been identified [3, 4], as causing a variety of clinical manifestations. Most of them are unique to a family (private) and therefore genotype-phenotype correlation is limited [5]. Diagnosing FD is challenging due to the range of disorders that mimic the disease and the great variety of atypical clinical presentations. As a result underdiagnosis and misdiagnosis of FD lead to late diagnosis, delays in appropriate treatment and a subsequent negative prognosis [6]. Human genetic analysis must be performed, in order to exclude or verify a mutation of the *GLA* [7]. Once a diagnosis has been made, biochemical and molecular genetic analysis, as well as genetic counselling, should be made available to all family members [8]. A detailed pedigree analysis for each patient

presenting with FD is crucial [9], as it can inform the diagnosis of the proband and the identification of all at-risk relatives [10].

Our aim is to report four novel *GLA* mutations resulting in FD and provide evidence of pathogenicity of the D313Y mutation regarding which contradictory data have been presented in the literature.

METHODS

Study design and setting

- This is a polycentric population screening study of individuals from Greece either demonstrating phenotypic traits suggestive for FD or belonging to families of patients with definite FD diagnosis and fulfilling anyone of the following inclusion criteria:
- Definite diagnosis of FD.
- Nephrological, cardiac or neurological symptoms suspicious of FD.
- Relatives of patients with definite FD diagnosis.

A cohort of 62 subjects from 19 unrelated families was involved in the study. Twenty-five of them were relatives of 9 patients with definite FD diagnosis and 18 were relatives of 10 individuals recruited as carriers of FD traits in whom a *GLA* mutation was detected. The presence of the D313Y mutation in Greek population was examined by genotyping of 145 apparently healthy subjects (70 males, 75 females). Written informed consent was obtained from each subject or an accompanying relative, where legally appropriate. The study was approved by the institutional review board of the University of Thessaly, Larissa.

Clinical assessment

Patients' medical records were revaluated and a detailed medical history of the family members was obtained especially in regard with heart or kidney disease, cerebrovascular events, death at young age and respective causes of death. All study participants underwent

physical examination particularly focused on cardiac, renal and neurological signs and symptoms. A detailed pedigree was constructed for every family and newborn screening was performed once.

Laboratory evaluation

In DBS, we measured α-Gal A activity in all male subjects by tandem mass spectrometry [11], lyso-Gb₃ in all subjects by HPLC and tandem spectrometry [12] as well as plasma and urine Gb₃ concentration in selected cases by tandem mass spectrometry [13].

Optical and electron microscopy was performed for the study of kidney biopsies occasionally.

Genotyping

Genomic DNA was extracted from peripheral blood using the iPrep Pure Link DNA blood kit (Invitrogen, Thermofisher, USA) according to manufacturer's instructions. All coding regions and exon-intron splice junctions of the *GLA* gene were analysed in a targeted custom next-generation sequencing (NGS) platform (Ampliseq custom panel, Thermo Scientific). Analysis of primary data was conducted with Ion Reporter software v.5.2 (Thermo Scientific). Common polymorphisms (UCSC Common SNPs) were excluded and pathogenicity of variations was predicted by bioinformatic analysis using PhyloP, SIFT, Grantham and PolyPhen tools, in comparison to their global (1000 Genomes Global Minor Allele Frequency, ExAC) and European frequency (5000 Exomes European Minor Allele Frequency). The characterization of variants was based on the recommendations of the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology [14].

Novel variants were verified by PCR in combination with Sanger sequencing.

Amplification of *GLA* exons (including exon-intron boundaries) was performed in 5 reactions

corresponding to exons 1, 2, 3, 4, and 5–7, using the primers included in Supplementary Material. A total of 100-200 ng of genomic DNA was amplified by PCR in a 30 □L reaction mixture using 200 \square M of each deoxynucleoside triphosphate, 30 pmol of each primer, 1.5 mM MgCl₂ and 1.0 U Taq polymerase (Invitrogen, Thermofisher, USA) in a 10x buffer supplied by the manufacturer. Reaction conditions were as following: For exons 1-3: 94 °C for 2 min, followed by 30 cycles of 30 sec at 94 °C, 30 s at 58 °C, 30 s at 72 °C, and a final extension at 72 °C for 5 min. For exons 4 and 5–7: 94 °C for 2 min, followed by 32 cycles of 30 sec at 94 °C, 30 sec at 54 °C, 30 sec for exon 4 or 75 sec for exons 5-7 at 72 °C, and a final extension at 72 °C for 5 min. All PCR reactions were carried out in the Veriti 96-Well Thermal Cycler (Applied Biosystems, Thermofisher, USA) PCR engine apparatus and the emerging PCR products were purified using the PureLink PCR Purification Kit system (ThermoFisher Scientific, USA). Sequencing was performed using the primers described in Table X, using a 3730 DNA Analyzer (Applied Biosystems, Thermofisher, USA) and BigDye Terminator DNA sequencing kit (Applied Biosystems, Thermofisher, USA) according to manufacturer's instructions.

RESULTS

Nine (all with definite FD) out of the 62 genotyped subjects were carrying five previously described *GLA* mutations: c.334C>T (p.Arg112Cys, R112C), c.644A>G (p.Asn215Ser, N215S), c.1153A>C (p.Thr385Pro, T385P), c.453C>G (p.Tyr151Ter, Y151X) and c.782G>T (p.Gly261Val, G261V).

The c.937G>T (p.Asp313Tyr, D313Y, NM_000169.2) mutation was revealed in seventeen individuals (54±14, range 27–78 years), seven males (61±11, range 45–78 years) and ten females (49±15, range 27–70 years) but in none of the healthy subjects. Patients' clinical and laboratory findings are presented in Table 1. All male patients (61±11, range 45–78 years) presented with an α-gal A activity decreased in a range of 56.2-87.5% comparing to

normal with the exception of one of them in whom α -gal A activity was within normal range. LysoGb₃ concentration was found normal (range 0.8-1.7 ng/mL) in all patients, while plasma and urine Gb₃ concentration was varying as shown in Table 1.

The diagnosis of FD was definitely posed according to the recently published criteria of the disease [15], in five carriers of this mutation (54 ± 9 , range 45-65 years). The first of them (patient no.1, Table 1), a 52-year-old man was initially diagnosed at the age of 48 with CKD stage III and was not presenting cardiac or other FD signs. Kidney biopsy performed because of non-nephrotic proteinuria, microscopic haematuria and raised serum creatinine, revealed focal and segmental glomerulosclerosis (FSGS) – collapsing variant. At that time enzyme activity and plasma lyso-Gb₃ concentration were normal. Three years after initial presentation the patient was suffering end stage renal disease and extreme acroparesthesias. Revaluation of kidney biopsy by higher magnification uncovered focal cytoplasmic microvacuolization of enlarged podocytes (Fig.1) while a decreased by 50% α -gal A activity and an increased plasma (7.52 nmol/mL, reference: 0.8-4.52) and urine Gb₃ concentration (147.49 nmol/g, reference: <29.00) were detected. The patient commenced dialysis and ERT with rapid clinical improvement.

A second carrier of the D313Y mutation was a 46-year-old female (patient no. 4, Table 1) who had suffered a TIA and two ischaemic strokes that had been considered of unknown origin. On revaluation, one year after the last stroke the patient appeared with microalbuminuria, oedema and acroparesthesias on both hands. The microalbuminuria was duplicated after three months and acroparesthesias worsened. Gb₃ concentration was pathological in urine (54.08 nmol/g) but normal in plasma. LysoGb₃ concentration was normal at that time and remained stable for one more year.

The mother of the above patient (patient no. 5, Table 1), 65 years old, had been diagnosed at the age of 50 with MS and was receiving relative medication, without clinical

benefit. She was also receiving treatment for pain in extremities that were attributed to RA. During the recent years the patient suffered mobility impairment (reported as spastic quadriplegia after a neurological examination), depression and dementia. The evaluation of the patient revealed pathological plasma Gb₃ concentration (4.7 nmol/mL), WML on brain MRI and normal kidney function. LysoGb₃ concentration was normal and remained so during a 6-month-follow up.

Another carrier of the D313Y mutation was a 45-year-old male (patient no.7, Table 1) on dialysis due to CKD by the age of 25. No kidney biopsy was performed at that time. He reports episodes of haematuria during childhood and adolescence, attributed at that time to vesicoureteral reflux. Enzyme activity was slightly decreased (2.4 µmol/l/h, reference: ≥2.6) and plasma lyso-Gb₃ concentration was normal. Brain MRI revealed WML and vertebrobasilar vessel changes. Moreover, increased echogenicity of cardiac interventricular septum on the echocardiogram and sensorineural hearing loss of higher frequencies. The patient commenced ERT.

The last patient with D313Y mutation definitely diagnosed as suffering FD was a 60-year-old female (patient no.17, Table 1) who was presenting cornea verticillata corneopathy, WML and ischaemic infarcts on brain MRI despite that no stroke is reported, acroparesthesias and GI symptoms (pain-diarrhoea) since adolescence, hypohidrosis, hearing loss and LVH.

Among the remaining twelve D313Y mutation carriers there were two patients with no FD signs and eight patients (55 ± 17 , range 27–78 years) presenting other FD signs mainly neurological that, however, cannot document a definite diagnosis of FD. Especially patient no.6, Table 1 (62-year-old female) suffers acroparesthesias and GI symptoms since adolescence and presents with lysoGb₃ at 1.7 ng/ml (reference: \geq 1.8). Accordingly patient no. 16, Table 1 (64-year-old male) presents with CKD, diabetes mellitus and hearing loss,

while on brain MRI presents multiple ischaemic infracts despite that no stroke is reported. The remaining two D313Y mutation carriers are apparently healthy.

Novel GLA mutations (4) (Fig. 2) were detected in thirteen members (39±18, range 1– 69 years) of four unrelated families (Table 2), all fulfilling the diagnostic criteria of a definite diagnosis of FD [15]. The c.835C>T mutation (p.Gln279Ter, Q279X) in exon 6 of the GLA gene was identified in four members (30±24, range 1–60 years) of a Greek family. The proband, a 31-year-old male, was presented at the age of 23 with proteinuria (3.5 gr/24 h), microscopic haematuria, slightly deteriorated kidney function, right bundle brunch block, mild hypertension and angiokeratomas in the arms and the loin area. He reported pain in the extremities especially during infections and inability to sweat. The α -gal A activity was close to zero. Cardiac MRI showed moderate LVH. The kidney biopsy showed cytoplasmic vacuolization and extended lysosomal accumulations in all types of kidney cells, especially in the podocytes (Fig. 3). The patient is currently under ERT with beneficial results in regard with kidney function, proteinuria and pain. Three other members of the family were revealed having with the same mutation and all presented clinical signs of FD. The proband's mother (60 years old) reported a possible TIA at the age of 32. Kidney examination showed albuminuria (> 500 mg/24 h), microscopic haematuria and normal kidney function. Cardiac MRI revealed severe LVH (cardiac interventricular septum over 19mm) and on skin examination she showed angiokeratomas in the arms. Her mother (the proband's grandmother) had died at the age of 62, due to cardiac arrest. She suffered from severe LVH and acroparesthesias, which at that time were attributed to Raynaud's phenomenon. The proband's sister (27-year-old) has elevated lyso-Gb₃ concentration (2.7 ng/ml) and screening of her newborn daughter revealed the mutation too.

Four patients (46±11, range 30–56 years) belonging to another Greek family were carrying the c.280T>A (p.Cys94Ser, C94S) mutation. The proband, a 48-year-old female, was

diagnosed with FD a year before after presenting increased lyso-Gb₃ concentration (5.4 ng/ml), increased plasma Gb₃ concentration (6.14 nmol/ml), microalbuminuria of no other origin, cornea verticillata corneopathy, acroparesthesias in both hands and dyshidrosis. Clinical and laboratory data of the other suffering members of the family are presented in Table 2.

The c.924A>C mutation (K308N - p.Lys308Asn) in exon 6 of the GLA gene was identified in three members (44±26, range 17–69 years) of a Greek family. The proband, a 46-year-old male, was presented at the age of 34 with albuminuria (0,5-0,6 gr/24h), microscopic haematuria since ten years and slight hypertension. The kidney function was normal. Kidney biopsy showed slight mesangial proliferative damages and cytoplasmic microvacuolization of podocytes. The α -gal A activity was almost zero. After 10-year-follow up he suffered of LVH and proteinuria (1.8 gr/24h). The patient is currently under ERT. His mother and daughter carry the same mutation. The probands grandmother had died at the age of 68, suffering of severe LVH and end-stage heart failure.

Lastly, the c.511G>A mutation (G171S - p.Gly171Ser) in exon 3 of the *GLA* gene was identified in two members (37 ± 4 , range 34-39 years) of an Albanian family living in Greece. The proband, a 39-year-old male, was diagnosed at the age of 32 with severely deteriorated kidney function and proteinuria. No biopsy was performed at that time, due to the small size of the kidneys. After nearly a year he presented with severe clinical and laboratory findings of acute renal failure and need of dialysis. Normal kidney function was never restored. FD was definitely diagnosed at the age of 37, as the α -gal A activity was extremely low and lyso-Gb₃ concentration was 11.9 ng/ml. The patient is suffering of LVH, increased pulmonary artery diameter, dilatation of the ascending aorta and aortic valve stenosis, because of which he underwent a valve replacement surgery. Ophthalmological evaluation indicated lipid deposition with blurriness of the cornea. The patient is currently under ERT. His 34-

year-old brother was identified with the same mutation and has extremely low α -gal A activity, lyso-Gb3 concentration 12.9 ng/ml and albuminuria of no other aetiology.

DISCUSSION

The D313Y mutation

Contradicting results about the pathogenicity of this mutation have been reported in the literature since its first description on 1993 [16]. The mutation has been detected in many series of patients presenting signs of FD [17, 18, 19, 20, 21, 22, 23, 24]. However, Niemann et al. [25] describes this variant as non pathogenic, although his two patients were presenting decreased α -gal A activity. Similarly, Oder et al. [26] supports that the D313Y genotype does not lead to severe organ manifestations as seen in genotypes known to be causal for classical FD and Froissart et al. [27] characterizes the mutation as "pseudodeficient allele" implying that it is a sequence variant which encodes an enzyme that is transported to the lysosomes, where it has about 75% of normal enzymatic activity. The D313Y mutation has been also referred as polymorphism [19], despite that, according to the ExAC and 1000 Genomes databases, its frequency in the World and European population is below 1%. Finally, it must be mentioned that by bioinformatics analysis this mutation is predicted as probably damaging (PolyPhen-2) or damaging (SIFT).

In our study five of seventeen carriers of the D313Y mutation (54 ± 9 , range 45-65 years) proved as suffering definite FD according to the recently published criteria of the disease [15]. The presentation of the disease in our patients indicates that the mutation results in a milder phenotype, with later onset of symptoms. This phenotype, also including milder mono- or oligosymptomatic cases [19], is characterised as atypical or type 2 [28]. The late onset of clinical symptoms and the milder than the typical phenotype of FD in these patients can be partly explained by the high α -gal A residual activity, since there is evidence that the

mutated α -gal A reaches intracellularly the lysosomes [29]. Actually, in all but one male D313Y carriers of our study α -gal A activity was decreased. Another reason for the decreased activity of the D313Y enzyme in plasma could be a functional intolerance to blood plasma neutral pH conditions. This effect is irreversible and, once in contact with a neutral or basic pH environment D313Y enzyme remains inactive, even if transferred to optimal pH [21, 29].

Interestingly, the vast majority of our D313Y patients were presenting with neurological symptoms and signs. Moreover, one of them was misdiagnosed as MS while two carriers of the mutation, in whom definite FD diagnosis has not been established as yet, are diagnosed and treated as MS. Similarly the detailed study of the renal biopsy in one of our cases underlies the significance of early detection of Fabry specific findings in cases with FSGS.

Novel mutations

The pathogenicity of the above mentioned novel mutations has been undoubtfully proved as all carriers suffer definite FD. Of them the c.835C>T (p.Gln279Ter, Q279X) is a nonsense mutation causing an interruption of the reading frame by a premature stop codon, which results in a truncated protein. A truncated protein and the subsequent loss of its functionality is strong evidence that the mutation is probably pathogenic for FD [14]. No amino acid change at this position has ever been described in NCBI and Fabry Database (http://fabry-database.org/) [30].

As far as the c.280T>A (C94S), c.924A>C (K308N), c.511G>A (G171S) mutations are considered, bioinformatics analysis supports their disease causing effect as they are predicted as probable damaging (PolyPhen-2) and/or damaging (SIFT). A further evidence of their pathogenicity could be the fact that the same aminoacid changes at the corresponding positions but in different nucleotides have already been described as disease causing [31, 32, 33].

CONCLUSIONS

We report four novel *GLA* mutations causing FD. Moreover, we offer strong evidence that the D313Y mutation could be pathogenic. It seems that this mutation is related with a later-onset milder than the typical phenotype with normal lysoGb₃ concentration.

Additionally our study underlines the significance of family members genotyping, newborn screening and genetic counselling in avoiding misdiagnoses and crucial delays of diagnosis and treatment of the disease. Finally we confirmed the fact that heterozygous females may develop mild to severe FD as well as that genotype-phenotype correlation does not exist even among the members of the same families.

Table 1. Characteristics of patients carrying the D313Y mutation. Plasma and urine Gb₃ concentration was measured only in patients 1, 4 and 5.

Patient /		Duariana
Gender /	FD related clinical findings	Previous
Age (years)		diagnosis
1 / M / 52	End stage renal disease – dialysis, acroparesthesias, renal	FSGS
	cysts, elevated plasma and urine Gb ₃ concentration	nephropathy
2 / F / 30	Healthy	no
3 / F / 70	Healthy	no
4 / F / 46	Three strokes of unknown origin, WML on brain MRI,	no
	micro-albuminuria, oedema, acroparesthesias, elevated urine	
	Gb ₃ concentration	
5 / F / 65	Multiple WML on brain MRI, depression, elevated plasma	MS, RA,
	Gb ₃ concentration	spastic
		quadriplegia
6 / F / 62	Acroparesthesias, GI symptoms since adolescence	no
7 / M / 45	End stage renal disease on dialysis, WML and vertebrobasilar	Nephropathy
	vessel changes on brain MRI, increased cardiac	of unknown
	interventricular septum echogenicity, hearing loss of higher	origin
	frequencies	
8 / M / 62	End stage renal disease on dialysis	no
9 / F / 36	WML on brain MRI, acroparesthesias	MS
10 / M / 78	End stage renal disease on dialysis	RA
11 / F / 47	LVH, CKD	no
12 / M / 68	No FD signs	Myopathy
13 / F / 46	No FD signs	NMO
14 / F / 27	WML on brain MRI	MS
15 / M / 61	End stage renal disease on dialysis	Diabetic
		nephropathy
16 / M / 63	CKD, hearing loss, multiple ischaemic infracts on brain MRI	no
17 / F / 59	Cornea verticillata, hearing loss, LVH, acroparesthesias, GI	no
	symptoms (pain – diarrhoea) since adolescence, hypohidrosis,	
	T2 -WML or ischaemic infracts on brain MRI	

Table 2. Clinical characteristics of patients with the five novel mutations.

Mutation (NM_000169.2)	Clinical data
c.835C>T	CKD, haematuria, proteinuria, pain in extremities, dyshidrosis,
p.Gln279Ter	angiokeratomas, zero activity of a-gal A, LVH, kidney biopsy consistent
Q279X	with FD, TIA, pathological elevated lyso-Gb ₃
c.280T>A	Microalbuminuria, end stage renal disease – dialysis, cornea verticillata
p.Cys94Ser	corneopathy, acroparesthesias, dyshidrosis, LVH, WML on brain MRI,
C94S	zero activity of a-gal A, pathological elevated lyso-Gb ₃ , pathological
	elevated Gb ₃ in plasma/urine
c.924A>C	CKD, haematuria, kidney biopsy findings related to FD, zero activity of
p.Lys308Asn	a-gal A, LVH
K308N	
c.511G>A	Proteinuria, end stage renal disease – dialysis, LVH, valvulopathy, cornea
p.Gly171Ser	verticillata corneopathy, extremely low activity of a-gal A, pathological
G171S	elevated lyso-Gb ₃

D313Y mutation of the GLA. (a) Glomerulus with segmental sclerosis - PAS x400. (b)
Segmental sclerosis with features of the "collapsing" variant (arrow) - Jones' silver x400. (c)
Pale appearing glomerulus with a small area of sclerosis adhering to Bowman's capsule
(arrow) - PAS x400. (d) Cytoplasmic microvacuolization of podocytes (arrow), suggestive
of FD - PAS x400.
Figure 2. Sanger confirmation of novel mutations.
Figure 3. Electron microscopy findings of renal biopsy from a male FD patient carrying the
Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding of FD, are
marked with black arrows in (a) methylene blue semithin section, (b) tubular cells and a
fibroblast and (c,d) podocytes.

Abbreviations
α -gal A: Enzyme α -galactosidase A; ACMG: American College of Medical Genetics and
Genomics; CKD: Chronic kidney disease; DBS: Dried blood spot; ERT: Enzyme
replacement therapy; FD: Fabry disease; FSGS: focal and segmental glomerulosclerosis;
GI: Gastrointestinal; GLA: α -galactosidase A gene; GL-3 or Gb3: globotriaosylceramide;
LVH: Left ventricular hypertrophy; lyso-Gb3: globotriaosylsphingosine; MRI: Magnetic
resonance imaging; MV: mean value; MS: Multiple sclerosis; NMO: Neuromyelitis optica;
RA: Rheumatoid arthritis; SD: Standard deviation; TIA: Transient ischemic attack; UT:
Under treatment; WML: White matter lesions.
Acknowledgements
We appreciate the help and advice from A. Andrikos, S. Kouzouka, A. Triantafillou, D.
Kaltsidou, A. Kaliantzoglou, A. Lysitska, K. Papadopoulos, K. Zoganas, G. Tsivgoulis, Z.
Tegou, A. Fountoglou, T. Karapanagiotidis, S. Patsialas.
Availability of data and materials
All data supporting our findings are included in the manuscript.
Authors' contributions
KK: collection, analysis and interpretation of the data, pedigree analysis, literature review,
drafting and revision of the manuscript. KS: Renal biopsy and analysis on electron
microscope. PP: Renal biopsy and analysis on optical microscope. MS, MZ and GL:
genotyping and bioinformatics analysis. KS, PP, EM, PK, CK, AO, JK: treating physicians of
patients. AEG: design and coordination of the study, revision of the manuscript. All authors
approved the final version of the manuscript.
Competing interests

KK received travel assistance from Shire and Genzyme and speaker's honoraria from Shire.
KS received travel assistance and speaker's honoraria from Shire. PP and AO received travel
assistance from Genzyme and Shire. CK received travel assistance from Shire. AEG received
research grants from Shire. The authors MZ, GL, MS, EM, PK, JK, report no competing
interests.
Funding
The study has been partially supported by a grant from the Research Committee of the
University of Thessaly.
Consent for publication

Written informed consents for this publication were obtained from patients. A copy of the consent form is available for review by the Editor of this journal.

Ethics approval and consent to participate

This report was approved by the institutional review board of the University of Thessaly,

Larissa. Written informed consents for participation were obtained from patients.

Page 20 of 32

361 References

- 1. Germain DP. Fabry disease. Orphanet J Rare Dis. 2010;5:30
- 2. Schiffmann R. Fabry disease. Pharmacol Ther. 2009;122:65-77
- 1000 Genomes Project Consortium, Abecasis GR, Auton A, Brooks LD, DePristo MA,
 Durbin RM, et al. An integrated map of genetic variation from 1,092 human genomes.
 Nature. 2012;491:56–65
- 4. The Human Gene Mutation Database at the Institute of Medical Genetics in Cardiff. http://www.hgmd.cf.ac.uk/. Accessed 13 June 2017
- 5. Laney DA, Fernhoff PM. Diagnosis of Fabry disease via analysis of family history. J Genet Couns. 2008;17:79–83
- 6. Mehta A, Ricci R, Widmer U, Dehout F, Garcia de Lorenzo A, Kampmann C, et al. Fabry disease defined: baseline clinical manifestations of 366 patients in the Fabry Outcome Survey. Eur J Clin Invest. 2004;34:236–42
- Gal A, Hughes DA, Winchester B. Towards a consensus in the laboratory diagnostics of Fabry disease – recommendations of a European expert group. J Inherit Metab Dis. 2011;34:509–14
- 8. Winchester B, Young E. Biochemical and genetic diagnosis of Fabry disease. In: Mehta A, Beck M, Sunder-Plassmann G, editors. Fabry disease: perspectives from 5 years of FOS. Oxford: Oxford PharmaGenesis; 2006. Chapter 18
- Hughes DA, Evans S, Milligan A, Richfield L, Mehta A. A multidisciplinary approach to the care of patients with Fabry disease. In: Mehta A, Beck M, Sunder-Plassmann G, editors. Fabry disease: Perspectives from 5 years of FOS. Oxford: Oxford PharmaGenesis; 2006. Chapter 35
- Laney DA, Fernhoff PM. Diagnosis of Fabry disease via analysis of family history. J Genet Couns. 2008;17:79–83

- 11. Chamoles NA, Blanco M, Gaggioli D. Fabry disease: enzymatic diagnosis in dried blood spots on filter paper. Clin Chim Acta. 2001;308:195-6
- 12. Aerts JM, Groener JE, Kuiper S, Donker-Koopman WE, Strijland A, Ottenhoff R, et al. Elevated globotriaosylsphingosine is a hallmark of Fabry disease. Proc Natl Acad Sci USA. 2008;105:2812–7
- 13. Boscaro F, Pieraccini G, la Marca G, Bartolucci G, Luceri C, Luceri F, et al. Rapid quantitation of globotriaosylceramide in human plasma and urine: a potential application for monitoring enzyme replacement therapy in Anderson-Fabry disease.

 Rapid Commun Mass Spectrom. 2002;16:1507-14
- 14. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17:405-24
- 15. Biegstraaten M, Arngrímsson R, Barbey F, Boks L, Cecchi F, Deegan P, et al. Recommendations for initiation and cessation of enzyme replacement therapy in patients with Fabry disease: the European Fabry Working Group consensus document. Orphanet J Rare Dis. 2015;10:36
- 16. Eng CM, Resnick-Silverman LA, Niehaus DJ, Astrin KH, Desnick RJ. Nature and frequency of mutations in the alpha-galactosidase A gene that cause Fabry disease. Am J Hum Genet. 1993;53:1186-97
- 17. Gaspar P, Herrera J, Rodrigues D, Cerezo S, Delgado R, Andrade CF, et al. Frequency of Fabry disease in male and female haemodialysis patients in Spain. BMC Med Genet. 2010;11:19
- 18. Wang RY, Bodamer OA, Watson MS, Wilcox WR; ACMG Work Group on Diagnostic Confirmation of Lysosomal Storage Diseases. Lysosomal storage diseases: diagnostic

- confirmation and management of presymptomatic individuals. Genet Med. 2011;13:457-84
- 19. Lukas J, Giese AK, Markoff A, Grittner U, Kolodny E, Mascher H, et al. Functional Characterisation of alpha-galactosidase a mutations as a basis for a new classification system in fabry disease. PLoS Genet. 2013;9:e1003632
- 20. Böttcher T, Rolfs A, Tanislav C, Bitsch A, Köhler W, Gaedeke J, et al. Fabry disease underestimated in the differential diagnosis of multiple sclerosis? PLoS One.
 2013;8:e71894
- 21. Lenders M, Duning T, Schelleckes M, Schmitz B, Stander S, Rolfs A, et al. Multifocal white matter lesions associated with the D313Y mutation of the a-galactosidase A gene. PLoS One. 2013;8:e55565
- 22. Samuelsson K, Kostulas K, Vrethem M, Rolfs A, Press R. Idiopathic small fiber neuropathy: phenotype, etiologies and the search for fabry Disease. J Clin Neurol. 2014;10:108-118
- Becker J, Rolfs A, Karabul N, Berlit P, Kraemer M. D313Y mutation in the differential diagnosis of white matter lesions: Experiences from a multiple sclerosis outpatient clinic. Mult Scler. 2016;22:1502-5
- 24. De Brabander I, Yperzeele L, Groote CC, Brouns R, Baker R, Belachew S, et al. Phenotypical Characterization of α -Galactosidase a Gene Mutations Identified in a Large Fabry Disease Screening Program in Stroke in the Young. Clin Neurol Neurosurg. 2012;115:1088-93
- 25. Niemann M, Rolfs A, Giese A, Mascher H, Breunig F., Ertl G, et al. Lyso-Gb3 indicates that the Alpha-Galactosidase A mutation D313Y is not clinically relevant for Fabry Disease. JIMD Rep. 2013;7:99-102

- 26. Oder D, Üceyler N, Liu D, Hu K, Petritsch B, Sommer C, et al. Organ manifestations and long-term outcome of Fabry disease in patients with the GLA haplotype D313Y. BMJ Open. 2016;6:e010422
- Froissart R, Guffon N, Vanier MT, Desnick RJ, Maire I. Fabry disease: D313Y is an alpha-galactosidase A sequence variant that causes pseudodeficient activity in plasma.
 Mol Genet Metab. 2003;80:307–14
- 28. Lenders M, Weidemann F, Kurschat C, Canaan-Kühl S, Duning T, Stypmann J, et al.

 Alpha-Galactosidase A p.A143T, a non-Fabry disease-causing variant. Orphanet J

 Rare Dis. 2016;11:54
- 29. Yasuda M, Shabbeer J, Benson SD, Maire I, Burnett RM, Desnick RJ. Fabry disease: characterization of alpha-galactosidase A double mutations and the D313Y plasma enzyme pseudodeficiency allele. Hum Mutat. 2003;22:486–92
- 30. Meiji Pharmaceutical university. http://fabry-database.org. Accessed 15 February 2017
- 31. Shabbeer J, Yasuda M, Benson SD, Desnick RJ. Fabry disease: Identification of 50 novel α -galactosidase mutations causing the classic phenotype and three-dimensional structural analysis of 29 missense mutations. Hum Genomics. 2006;2:297-309
- 32. Blaydon D, Hill J, Winchester B. Fabry disease: 20 novel GLA mutations in 35 families. Hum Mutat. 2001;18:459
- 33. Shimotori M, Maruyama H, Nakamura G, Suyama T, Sakamoto F, Itoh M, et al. Novel mutations of the GLA gene in Japanese patients with Fabry disease and their functional characterization by active site specific chaperone. Hum Mutat. 2008;29:331

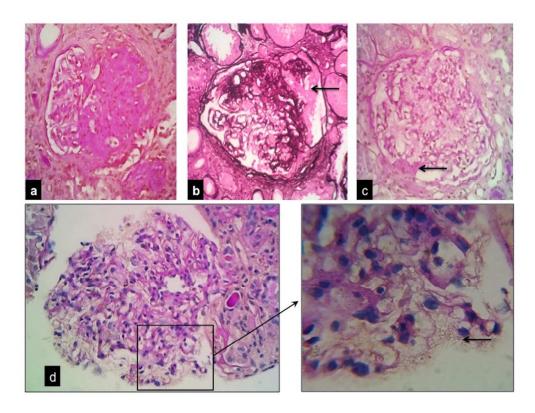


Figure 1. Optical microscopy findings of renal biopsy from a male FD patient carrying the D313Y mutation of the GLA. (a) Glomerulus with segmental sclerosis - PAS x400. (b) Segmental sclerosis with features of the "collapsing" variant (arrow) - Jones' silver x400. (c) Pale appearing glomerulus with a small area of sclerosis adhering to Bowman's capsule (arrow) - PAS x400. (d) Cytoplasmic microvacuolization of podocytes (arrow), suggestive of FD - PAS x400.

254x190mm (300 x 300 DPI)

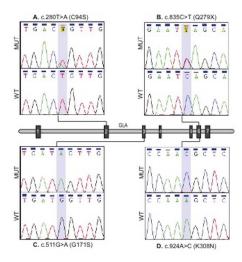


Figure 2. Sanger confirmation of novel mutations.

254x142mm (300 x 300 DPI)

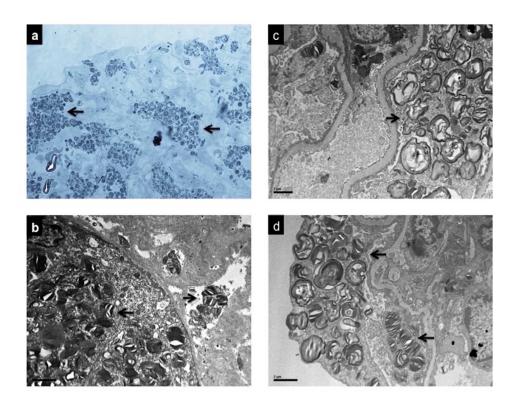


Figure 3. Electron microscopy findings of renal biopsy from a male FD patient carrying the Q279X mutation. Multi-lamellated myelin figures ("zebra" bodies), typical finding of FD, are marked with black arrows in (a) methylene blue semithin section, (b) tubular cells and a fibroblast and (c,d) podocytes.

254x190mm (300 x 300 DPI)

Supplementary file: Primer sequences for amplification and sequencing of the GLA gene.

Fragment	Sequence	Size (bp)
Genomic DNA amplification		
Exon 1	Forward: 5'- CCCAGTTGCCAGAGAAACAATAAC-3'	391
	Reverse: 5'-AGACTCTCCAGTTCCCCAAACAC -3'	
Exon 2	Forward: 5'-CCCAAGGTGCCTAATAAATGG -3'	337
	Reverse: 5'-CCATGAGGGCTGTTTCTAA -3'	
Exon 3	Forward: 5'-CGCAGCCTGGAATGGTTCTCTC -3'	323
	Reverse: 5'-CTCAGCTACCATGGCCTCA -3'	
Exon 4	Forward: 5'-AGCTGGAAATTCATTTCTTT -3'	212
	Reverse: 5'-TTGGTTTCCTTTGTTGTCA -3'	
Exons 5-7	Forward: 5'-AAACTCAAGAGAAGGCTACAAGT -3'	1280
	Reverse: 5'-AAAAAGGTGGACAGGAAGTAGTAGT 3'	
Sequencing Primers		
Exon 1 forward	5'- CCCAGTTGCCAGAGAGAAACAATAAC-3'	
Exon 2 forward	5'-CCCAAGGTGCCTAATAAATGG -3'	
Exon 3 forward	5'-CGCAGCCTGGAATGGTTCTCTC -3'	
Exon 4 forward	5'-TTGGTTTCCTTTGTTGTCA -3'	
Exons 5-6 forward	5'-AAACTCAAGAGAAGGCTACAAGT -3'	
Exons 7 forward	5'-TGAATGCCAAACTAACAGG	

Koulousios et al. STROBE Research Checklist

	Item No	
Title and abstract	1	Title [Page 1]: Fabry disease due to D313Y and novel GLA mutations
		Abstract [Page 3]
		The study is a polycentric observational cohort study which addresses the question of
		pathogenicity of specific Fabry disease mutations, using cohort data.
		Abstract [Page 3]: The abstract summarises the objectives, settings and participants primary and secondary outcome measures, results and conclusions of the study.
Introduction Dealerment/rationals	2	Introduction [Dage 4]
Background/rationale	2	Introduction [Page 4] Introduction to the scientific background of Fabry disease and <i>GLA</i> mutations.
Objectives	3	Introduction [Page 5]
		Aim of the study is to report five novel GLA mutations resulting in FD and provide
		evidence of pathogenicity of the D313Y mutation regarding which contradictory data
		have been presented in the literature.
Methods		
Study design	4	Methods - Study design and setting [Page 5]
		- Polycentric observational cohort study, multiple patient cases from various medical
		units all over Greece (polycentric study).
		- The study was approved by the institutional review board of the University of
		Thessaly, Larissa.
Setting	5	Methods - Study design and setting, Clinical assessment [Page 5]
		- Patient cases from multiple medical units in various locations all over Greece.
		- Family members of the majority of the Greek FD patients, who were diagnosed
		during the last years.
		- Pedigree analysis after personal interviews with the selected individuals in their
		places of residence.
Participants	6	Methods - Study design and setting, Clinical assessment [Page 5]
		Participants of the study fulfilled one of the following eligibility criteria:
		i) relatives of a patient with definite FD diagnosis.
		ii) patient cases with nephrological, cardiac or neurological symptoms suspicious of
		FD diagnosis from various medical units all over Greece.
		- Additional genotyping of 145 apparently healthy subjects.
		- The source of participants selection was their medical records (relation with a FD
		patient, patients with FD related symptoms), analysed in collaboration with their
		treating physicians.
		- Patients follow-up in collaboration with their treating physicians. Revaluation of the
		clinical status of each selected individual.
Variables	7	Clinical assessment - Laboratory evaluation - Genotyping - [Pages 5-6]

57

58 59

60

Variables used in the study:

- i) Heart or kidney disease, cerebrovascular events, death at young age and respective causes of death.
- ii) FD related cardiac, renal and neurological signs and symptoms.
- iii) Familiar relationship to a FD patient.
- iv) Enzyme α -Gal A activity in all male subjects.

BMJ Open

- v) Lyso-Gb₃ in all subjects.
- vi) Plasma and urine Gb₃ concentration in selected cases.
- vii) Presence of a GLA mutation.
- viii) Verification of novel mutations.
- ix) Evaluation of kidney biopsies.

Diagnostic criteria [Page 6]

The diagnosis of FD was posed according to the recently published criteria of the disease [Biegstraaten et al. 2015]

Data sources/ measurement

Clinical assessment - Laboratory evaluation - Genotyping - page 5-6

Method of assessment for each variable:

- i) Medical history.
- ii) Physical examination.
- iii) Pedigree analysis.
- iv) In DBS by tandem mass spectrometry.
- v) In DBS by HPLC and tandem spectrometry.
- vi) Tandem mass spectrometry.
- vii) Next-generation sequencing.
- viii) Standard Sanger sequencing using Variant Reporter software v1.1 (Applied Biosystems).
- ix) Optical and electron microscopy.

Bias

9 Study design and setting [Page 5]

Cross examination for the presence of D313Y mutation by genotyping of 145 apparently healthy subjects.

Laboratory evaluation [Page 6]

Measurement of α -Gal A activity only in male subjects.

Genotyping [Page 6]

10

Exclusion of common polymorphisms (UCSC Common SNPs) by bioinformatic analysis using PhyloP, SIFT, Grantham and PolyPhen tools, in comparison to their global (1000 Genomes Global Minor Allele Frequency, ExAC) and European frequency (5000 Exomes European Minor Allele Frequency).

Study size

Study design and setting [Page 5]

A cohort of 62 subjects from 19 unrelated families was involved in the study. Twenty-five of them were relatives of 9 patients with definite FD diagnosis and 18 were relatives of 10 individuals recruited as carriers of FD traits in whom a GLA mutation was detected.

Quantitative variables

11 Laboratory evaluation [Page 6]

In DBS, α -Gal A activity was measured in all male subjects by tandem mass spectrometry, lyso-Gb₃ in all subjects by HPLC and tandem spectrometry as well as

plasma and urine Gb₃ concentration in selected cases by tandem mass spectrometry.

Statistical methods

Age of each patient or patient group is presented either as an absolute number or as mean value \pm the Standard Deviation (mean \pm SD).

Results

Participants

13 Results [Page 7]

62 subjects were potentially eligible for participation in the study and were genotyped for a *GLA* mutation, clinically analysed and under follow-up for a significant period of time.

Abstract [page 3]

The 62 subjects were selected between 3 categories of patient cases:

- i) Family members of unrelated patients with definite FD diagnosis.
- ii) Clinically suspected cases.
- iii) Family members of the previous category.

Descriptive data

4 Abstract [Page 3]

- i) 25 family members of 9 unrelated patients with definite FD diagnosis.
- ii) Ten clinically suspected cases (nephrological, cardiac or neurological symptoms suspicious of FD diagnosis) from various medical units all over Greece.
- iii) 18 family members of the previous category.

Results [Pages 6-11]

- i) Patients carrying the D313Y mutation: 17 (54±14, range 27–78 years). [page 6]
- ii) Male patients carrying the D313Y mutation: 7 (61±11, range 45–78 years). [page 6]
- iii) Female patients carrying the D313Y mutation: 10 (49±15, range 27–70 years). [page 6]
- iv) Patients carrying novel *GLA* mutations: 16 members (42±19, range 1–73 years) of 5 unrelated families, all fulfilling the diagnostic criteria of a definite diagnosis of FD. [page 8]
- v) Patients identified with the novel c.835C>T mutation: 4 members (30±24, range 1–60 years)
- of a Greek family. Proband of the family: a 31-year-old male with typical FD clinical and laboratory phenotype. [page 8,9]
- vi) Patients identified with the novel c.280T>A mutation: 4 members (46±11, range 30–56 years) belonging to another Greek family. Proband of the family: a 48-year-old female with typical FD clinical and laboratory phenotype. [page 9]
- vii) Patients identified with the novel c.924A>C mutation: 3 members (44±26, range 17–69 years) of a Greek family. Proband of the family: a 46-year-old male with typical FD clinical and laboratory phenotype. [page 9,10]
- viii) Patients identified with the novel c.511G>A mutation: 2 members $(37 \pm 4, \text{ range } 34-39 \text{ years})$ of an Albanian family living in Greece. Proband of the family: a 39-year-old male with typical FD clinical and laboratory phenotype. [page 10]

Outcome data

Results [pages 6-11]

- i) Six (all with definite FD) out of the 62 genotyped subjects were carrying four previously described *GLA* mutations. **[page 6]**
- ii) The D313Y mutation was revealed in 17 individuals, 7 males and 10 females. [page 6]
- iii) The novel c.835C>T, c.280T>A, c.924A>C and c.511G>A *GLA* mutations were detected in 13 members of 4 unrelated families.

Main results

16 Results [pages 6-11]

i) Six (all with definite FD) out of the 62 genotyped subjects were carrying four previously

described GLA mutations. [page 6]

- ii) The D313Y mutation was revealed in 17 individuals, 7 males and 10 females. [page 6]
- iii) The D313Y mutation was revealed in none of the healthy subjects. [page 6]
- iv) The diagnosis of FD was definitely posed in 5 carriers of the D313Y mutation. [page 6-8]
- v) Two D313Y mutation carriers were patients presenting no FD related signs or symptoms.

[page 8]

- vi) Eight D313Y mutation carriers were patients presenting other FD signs mainly neurological that, however, can not document a definite diagnosis of FD. [page 8]
- vii) Two D313Y mutation carriers were apparently healthy. [page 8]
- viii) The diagnosis of FD was definitely posed in all carriers of the novel mutations reported.

[page 8-11]

Other analyses

- i) Bioinformatics analysis of every GLA mutation reported, by (PolyPhen-2) or (SIFT) systems.

 [page 12-13]
 - ii) Novel mutations verification by standard Sanger sequencing using Variant Reporter software v1.1 (Applied Biosystems). [page 6]

Discussion

Key results

- 8 i) Four novel GLA mutations causing classical Fabry Disease are reported. [page 14]
 - ii) Strong evidence that the D313Y mutation could be pathogenic is offered. [page 14]

Limitations

- i) The main limitation is the lack of detailed clinical data in older participants. [page 4]
- ii) Biopsies of affected organs, the gold standard of definite diagnosis, are not available in all cases. [page 4]

Interpretation

Discussion [Pages 11-13]

- Five of seventeen carriers of the D313Y mutation proved as suffering definite FD according to the recently published criteria of the disease. The presentation of the disease in our patients indicates that the mutation results in a milder phenotype, with later onset of symptoms. [page 12]
- Contradicting results about the pathogenicity of this mutation have been reported in the literature since its first description on 1993. [page 12]
- The mutation has been detected in many series of patients presenting signs of FD. However, Niemann et al. describes this variant as non pathogenic, although his two patients were presenting decreased α-gal A activity. Similarly, Oder et al. supports that the D313Y genotype does not lead to severe organ manifestations as seen in genotypes known to be causal for classical FD and Froissart et al. [i] characterizes the mutation as "pseudodeficient allele" implying that it is a sequence variant which encodes an enzyme that is transported to the lysosomes, where it has about 75% of normal enzymatic activity. The D313Y mutation has been also referred as polymorphism. [page 12]
- The vast majority of the D313Y patients of the study were presenting with neurological symptoms and signs. [page 12]
- Kidney biopsy is significant for a FD diagnosis in patients with renal manifestations of the disease [page 7, 9-11]
- The pathogenicity of the four novel mutations reported has been undoubtfully proved as all carriers suffer definite FD. [page 13]

Generalisability 21

1 Discussion [Pages 12-13]

Conclusion [Page 13-14]

The study underlines the significance of:

- i) Diagnosis of FD and possible treatment of patients carrying the 5 novel and the D313Y *GLA* mutations.
- ii) Family members genotyping.
- iii) Newborn screening and genetic counselling.
- iv) Avoiding misdiagnoses and crucial delays of diagnosis and treatment of the disease.

The study confirmed the facts that:

- i) Heterozygous females may develop mild to severe FD.
- ii) Misdiagnosis is common in FD.
- iii) Genotype-phenotype correlation does not exist even among the members of the same families.

Other information

Funding

22 Funding [Page 19]

The study has been partially supported by a grant from the Research Committee of the University of Thessaly.